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Isolation and Identification of Endophytic Fungi from Leaves, Stem and Flower of *Clerodendrum thomsoniae Balf. f.*

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Abstract: ***Endophytic fungi are those living inside the host plant without causing any apparent negative effect on the host plant. 117 isolates of endophytic fungi were isolated from the leaves, stem and flower of Clerodendrum thomsoniae. Collected from different areas of Mysore. All isolates were identified based on characteristics of the isolates such as colony appearance, mycelial texture, conidial spores size, shape, pigmentation, and other morphological characteristics observed using a microscope. The isolates of endophytic fungi were identified are Ascomycota and Basidiomycota and classified genera Sterile hyphae, Lasiodiplodiae sp, Rhizopus stolonifera sp, Aspergillus sp, Aspergillus conidiophore, Mycelium of Lentinus, Nigrospora sp, Nigrospora zimmermaniae sp, Curvularia sp, Collectrotrichum sp, Aspergillus fumigates sp. The overall colonization rate of endophytes in leaves, stem and flower were found to be 96.66%. The leaf showed a low percentage of colonization frequency of the endophytic fungi when compared to stems and flower segments.***

Keywords: Endophytic fungi, Clerodendrum thomsoniae, isolation, and identification.

I. INTRODUCTION

Endophytes are micro-organisms that colonize plant tissues for at least part of their life cycle without causing disease symptoms in their host [1]. Plants may serve as a reservoir of the large number of micro-organisms known as endophytes. Endophytic fungi also called plant-hidden fungi are, defined as fungi that have part or all of their life cycle living naturally in the plant tissues without causing disease symptoms. Endophyte microbes are microbes that live inside plant tissues. Endophytic fungi are microorganisms most commonly found in plants [2], [3]. Endophytic fungi can be isolated from the leaves, stems, flowers, fruits, and seeds [4]. Endophytic fungi are one of the interesting microbial symbionts with a plant that gives benefits their host's defense from natural enemies [5]. Some endophytic fungi have produced toxic substances capable of protecting host plants against phytopathogens, insects, nematodes, and herbivores [6]. Endophytic fungi can become a saprophyte when host plant [7]. The relationship between the endophytic and the host is symbiont mutualism which both gain benefits each other to survive. In this study, we used *Clerodendrum thomsoniae*. This plant is a member of the family Fabaceae, Lamiaceae. *C. thomsoniae* is known to be a potential pharmacological plant because of its ability to produce secondary metabolites. Other members of the genus are reported to be used medicinally in India, China, Thailand and Japan for the treatment of such diseases as syphilis, typhoid, cancer, jaundice and hypertension (Shrivastav and Patel [8]. The purpose of this study is to identify the endophytic fungi from the leaves and stems of plants *C.thomsoniae* based on morphological characteristics as a preliminary study to determine the diversity of fungi found in the leaves and stems of plants *C. thomsoniae*.

II. MATERIALS AND METHODS

A. Isolation and culturing of endophytic Fungi

The fresh leaves, flowers, and stem parts were used for the isolation of endophytic fungi. The plant samples were stored in sealed plastic bags at 4°C until processed. Ripe, healthy leaves and stems of *C. thomsoniae* were washed thoroughly under running tap water, then the samples were sterilized by dipping them in 75% ethanol for 30 s, followed by immersing in 3 % sodium hypochlorite several times, rinsed in sterile distilled water, and finally dried on sterile filter paper on a petri dish. A piece of each leaf and stem was removed with a sterile scalpel and then cut into small pieces about 1 to 1.5 cm, each piece was put on a Petri plate containing Potato Dextrose Agar (PDA) medium and incubated at room temperature (27-28 ̊C) to promote fungal growth and sporulation. After 7-8 days, individual hyphal tips of actively growing fungi were picked up for subculturing by inoculating them onto a new PDA medium plate individually and incubated at room temperature (27 ̊C) for one week. The purified fungal isolates were labeled for further use.

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B. Morphological Identification

Identification of endophytic fungal isolates was done by observing cotton blue stained slides prepared from stock cultures using a bright-field and phase contrast microscope. Identification was based on morphological characteristics such as growth pattern, hyphae, the color of the colony, surface texture, marginal character, aerial mycelium, mechanism of spore production, and characteristics of conidia. Obtained data were then compared with the descriptions of endophytic fungi and identified based on [9], [10].

C. Statistical Analysis

Colonization frequency (CF)

To know the endophyte richness, the frequency of fungal endophytes harbored in plant species was calculated by the number of segments colonized by endophyte species divided by the total number of segments examined × 100.

$$CF\% = \frac{\text{No. of individual fungi recorded}}{\text{Total no. of segments examined}} \times 100$$

III. RESULTS

About 121 segments (40 segments leaves and flower, 41 segments stems) of the *C. thomaoniae* plant were screened for isolation of the endophytic fungi. In this study, a total of 117 endophytes were isolated. The endophytes were isolated using potato dextrose agar (PDA). The total numbers of colonies of endophytic fungi from the leaves were lower than the ones from the stems.

Based on morphological characteristics identified species were (Figures 1, 2, and 3) Sterile hyphae., *Lasiodiplodia* sp., *Rhizopus stolonifera* sp., unidentified., *Aspergillus* sp., *Aspergillus* conidiophore sp., *Corynespora cassiicola* sp., Mycelium of *Lentinus*. From leaves samples. From flower samples, fungi were Sterile hyphae., *Candida albicancs* sp., *Nigrospora vesicularis* sp., *Curvularia australiensis* sp., *Rosellinia necatrix* sp., *Nigrospora bumbusae* sp., *Fusarium avenaceum* sp., *Nigrospora* sp. From stem samples, the fungi were *Paecilomyces* sp., *Collectrotrichum* sp., *Arthrinium* sp., *Penicillium* sp., *Epicoccum nigrum* sp., *Aspergillus niger* sp., *Aspergillus flavus*., *Aspergillus fumigates*., *Rhizoctonia solani* sp. The results obtained were then compared with the literature and monographs. Based on the results of isolation from leaves, flowers and stems of *clerodendrum thomsoniae*, 117 isolates were obtained. The number of isolates from the leaves was 38 isolates, while from flowers was 39 isolates and stems was 40 isolates.

Table 1: Endophytic fungi isolated from the *C. thomsoniae* plant.

Site of location	No. of samples	No. of fungi isolated	CF (%)
Leaves	40	38	95.00
Flower	40	39	97.50
Stem	41	40	97.50
Total	121	117	96.66

C. thomsoniae: Clerodendrum thomsoniae, CF: Colonization frequency.

A. Identification of endophytic fungi from leaves of Clerodendrum thomsoniae

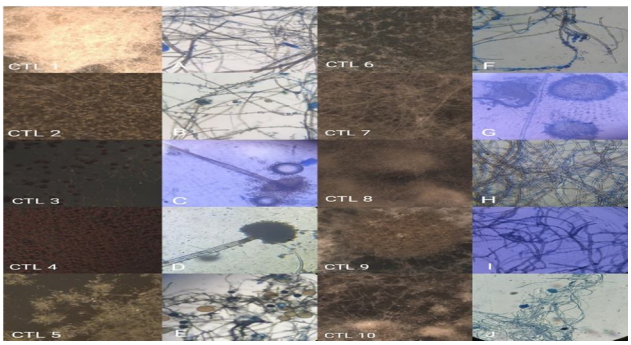


Figure 1. Light microscopic photographs of endophytic fungi from *Clerodendrum thomsoniae* leaves. (CTL 1-10 is Stereo microscopic photographs). A: Sterile hyphae., B: *Lasiodiplodiae* sp., C: *Rhizopus stolanifer* sp., D: *Aspergillus* sp., E: Unidentified., F: Unidentified., G*: Aspergillus* conidiophore sp., H: Sterile hyphae., I: *Corynespora cassiicola* sp., J : Mycelium of *Lentinus.*

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Table 2.1: Endophytic fungi isolated from the leaves of *Clerodendrum thomsoniae*

Sl.No	Isolated endophytes	Phylum	No. of segments	Total (%)
1	Sterile hyphae	-	8	20.00
2	*Lasiodiplodiae* sp.	Ascomycota	2	5.00
3	*Rhizopus stolonifera* sp.	Zygomycota	3	7.50
4	*Aspergillus* sp.	Ascomycota	11	27.50
5	Unidentified	-	2	5.00
6	Unidentified	-	1	2.50
7	*Aspergillus* conidiophore sp.	Ascomycota	2	5.00
8	Sterile hyphae	-	3	7.50
9	*Dermatophyte* sp.	Ascomycota	2	5.00
10	Mycelium of *Lentinus*	Basidiomycota	4	10.00
Total				38(95.00)

B. Identification of endophytic fungi from flower of Clerodendrum thomsoniae

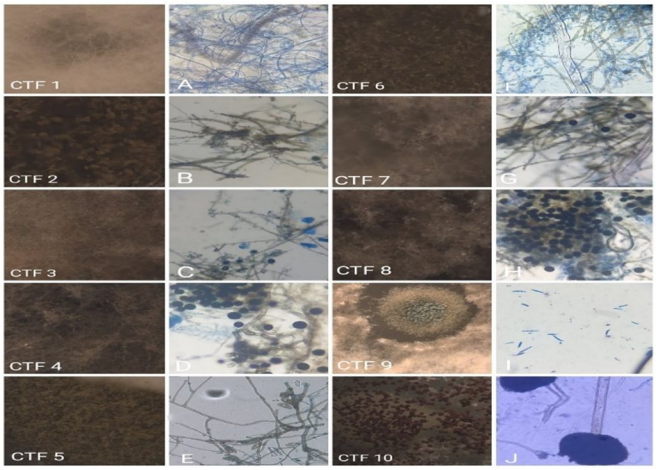


Figure 2: Light microscopic photographs of endophytic fungi from *Clerodendrum thomsoniae* flowers (CTF 1-10 Is Stereo photographs). A: Sterile hyphae., B: *Candida albicans* sp., C: *Nigrospora zimmermanie* sp., D: *Nigrospora vesicularis* sp., E: *Curvularia australiensis* sp., F: *Rosellinia necatrix* sp., G: *Nigrospora bumbuusae* sp., H*: Nigrospora* sp., I: *Fusarium avenaceum* sp., J: *Aspergillus* sp.

Table 2.2: Endophytic fungi isolated from the flower of *Clerodendrum thomsoniae*

Sl.No	Isolated endophytes	Phylum	No. of segments	Total(%)
1	Sterile hyphae	-	4	10.00
2	*Candida albicans* sp.	Ascomycota	2	5.00
3	*Nigrospora zimmermanie* sp.	Sordariomycetes	2	5.00
4	*Nigrospora vesicularis* sp.	Sordariomycetes	3	7.50
5	*Curvularia australiensis* sp.	Ascomycota	8	20.00
6	*Rosellinia necatrix* sp.	Ascomycota	2	5.00
7	*Nigrospora bumbusae* sp.	Sordariomycetes	4	10.00
8	*Nigrospora* sp.	Sordariomycetes	7	17.50
9	*Fusarium avenaceum* sp.	Ascomycota	4	10.00
10	*Aspergillus* sp.	Ascomycota	3	7.50
Total				39(97.50)

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C. Identification of endophytic fungi from Stem of Clerodendrum thomsoniae

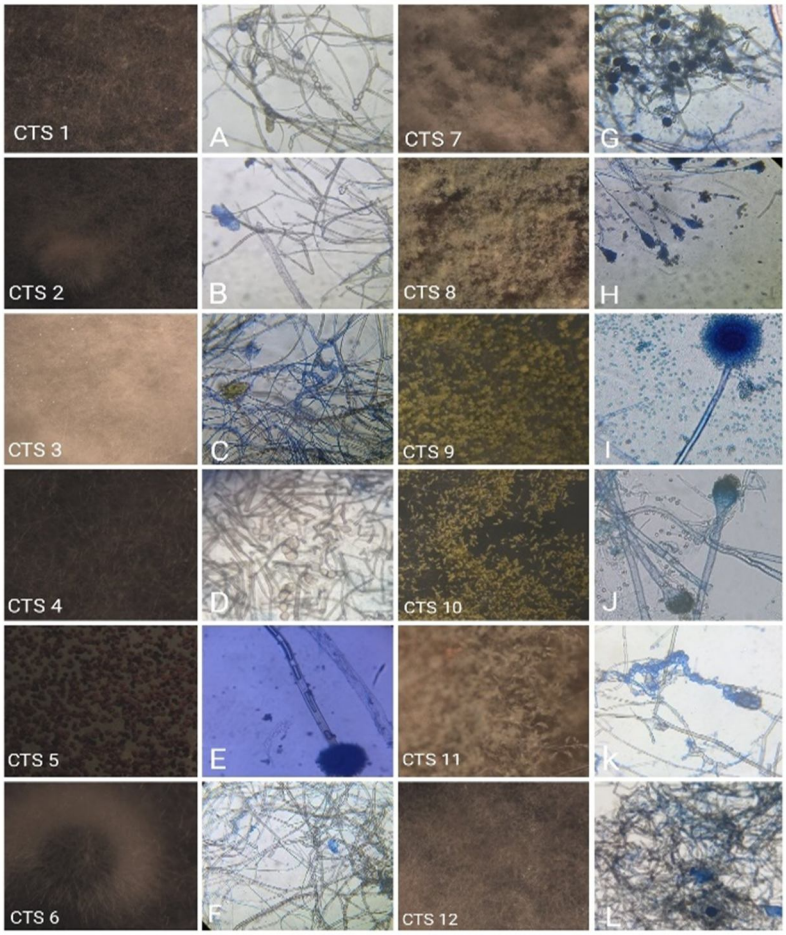


Figure 3: Light microscopic photographs of endophytic fungi from *Clerodendrum thomsoniae* stems (CTS 1-10 is Stereo microscopic photographs). A: *Paecilomyces* sp., B: *Collectrotrichum* sp., C: *Cylindrocladium* sp., D: *Arthrinium* sp., E: *Aspergillus niger* sp., F: *Epicoccum nigrum* sp., G: *Nigrospora* conidia sp., H: *Penicillium* sp., I: *Aspergillus flavus.*, J: *Aspergillus fumigates.*, K: *Ampelomyces* sp., L: *Rhizoctonia solani* sp.

Table 2.3: Endophytic fungi isolated from the stem of *Clerodendrum thomsoniae.*

Sl.No	Isolated endophytes	Phylum	No. of segments	Total (%)
1	*Paecilomyces* sp.	Ascomycota	2	4.87
2	*Collectrotrichum* sp.	Ascomycota	2	4.87
3	*Cylindrocladium* sp.	Ascomycota	2	4.87
4	*Arthrinium* sp.	Ascomycota	2	4.87
5	*Aspergillus* sp	Ascomycota	8	19.10
6	*Epicoccum nigrum* sp.	Ascomycota	3	7.31
7	*Nigrospora* conidia	Sordariomycetes	4	9.75
8	*Penicillium* sp.	Ascomycota	5	12.13
9	*Aspergillus flavus* sp.	Ascomycota	3	7.31
10	*Aspergillus fumigates* sp.	Ascomycota	4	9.75
11	*Ampelomyces* sp.	Ascomycota	2	4.87
12	*Rhizoctonia solani* sp.	Basidiomycota	3	7.31
Total				40(97.50)

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IV. DISCUSSION

Identification is based on the morphological characteristics of the fungi growing on the culture medium (PDA). The morphology includes macroscopic and microscopic characteristics. The macroscopic identification of colonies such as color diameter, colony growth, and colony reverse, observation was done for seven days during the fungal culturing.

Microscopic characterization was done by observing the shape and size of conidia, and hyphae. Observation of conidia, including its arrangements (singular, chain or cluster) number of cells (unicellular or multicellular) and measurement of conidia. Observation of hyphae was also performed on the presence or absence of septa in hypha, its shape, morphology, and modification of hyphae.

The results obtained were then compared with the literature and monographs. Based on the results of isolation from leaves, flowers and stems of *clerodendrum thomsoniae*, 117 isolates were obtained. The number of isolates from the leaves was 38 isolates, while from flowers was 39 isolates and from stems was 40 isolates.

Identification of fungi can be done conventionally by observing the morphological characters and comparing them to descriptions from the literature or monograph. The morphological characteristics which are observed include microscopic and macroscopic characteristics. Macroscopic observation based on the characteristics. Macroscopic observation is based on the characteristics of the fungal colony on agar media like color, texture, reverse side, and margins [11].

Macroscopic and microscopic characteristics of fungal isolates can be observed in the presence or absence of conidia, conidia shape, conidia arrangement, and size, conidiophore, hyphae (septum or singular), and hypha's pigment. But the microscopic observation only the teleomorph is found then the fungi classified in the Ascomycota. When the teleomorph of such fungi was found, it is classified in the Ascomycota under a different name. Both types of conidia (teleomorph and anamorph) are not found, then the fungi are placed in the sterile hyphae group [12], [13], [14].

The result showed that most of the fungal isolates were purified belonging to Ascomycota fungi and Basidiomycota fungi. A large number of these members did not show sexual structure so difficult to determine their class. The result also discovered 8 fungal isolates that did not show the formation of conidia. These isolates belonged to the mycelia sterile group because they did not show the form of anamorph or teleomorph during the observation process. Fungi isolates that were included in the mycelia sterile group were *Rhizoctonia* sp., and other fungi isolates that can not be identified because they present only a set of hyphae.

The presence of endophytic fungi in a plant depends on the species of plant, environment and isolation methods. The amount of endophytic fungi isolates that are found depends on the diversity and distribution of endophytic fungi exit in the host plants. Each part of the plant will give a different number of endophytic fungal isolates. The most dominant endophytic fungi present inside plant tissues indicated that the fungi were the most widely distributed species in the plant. This could be known by the percentage of endophytic fungi that have been found from the isolation process.

A more realistic approach is needed to characterize the endophyte species from a single host or group of hosts. The number of samples required for the isolation of the endophytic fungus related to the distribution and abundance of fungi in the host plant and the tissue types (stem, root, bark). A more intensive sampling method will increase the recovery of rare species, which are likely also to occur on many hosts, but the most common species on a specific host will be widely distributed on that host [15].

V. CONCLUSION

Endophytic organisms have received considerable attention as they are found to protect their hosts against pests, and pathogens. In this study, a total of 117 endophytes were isolated from the *C. thomsoniae*, a well-known medicinal plant that contains various chemical compounds and its identification based on morphological characteristics, of which 69 (57.02%) are Ascomycota, seven are Basidiomycota (5.78%), twenty are Sordariomycetus (16.52%) and three are Zygomycota (2.47%), two isolates are unidentified sp., fifteen isolates are Sterile hyphae (12.39%) and four isolates are mycelia of *Lentinus* (3.30%).

VI. ACKNOWLEDGMENT

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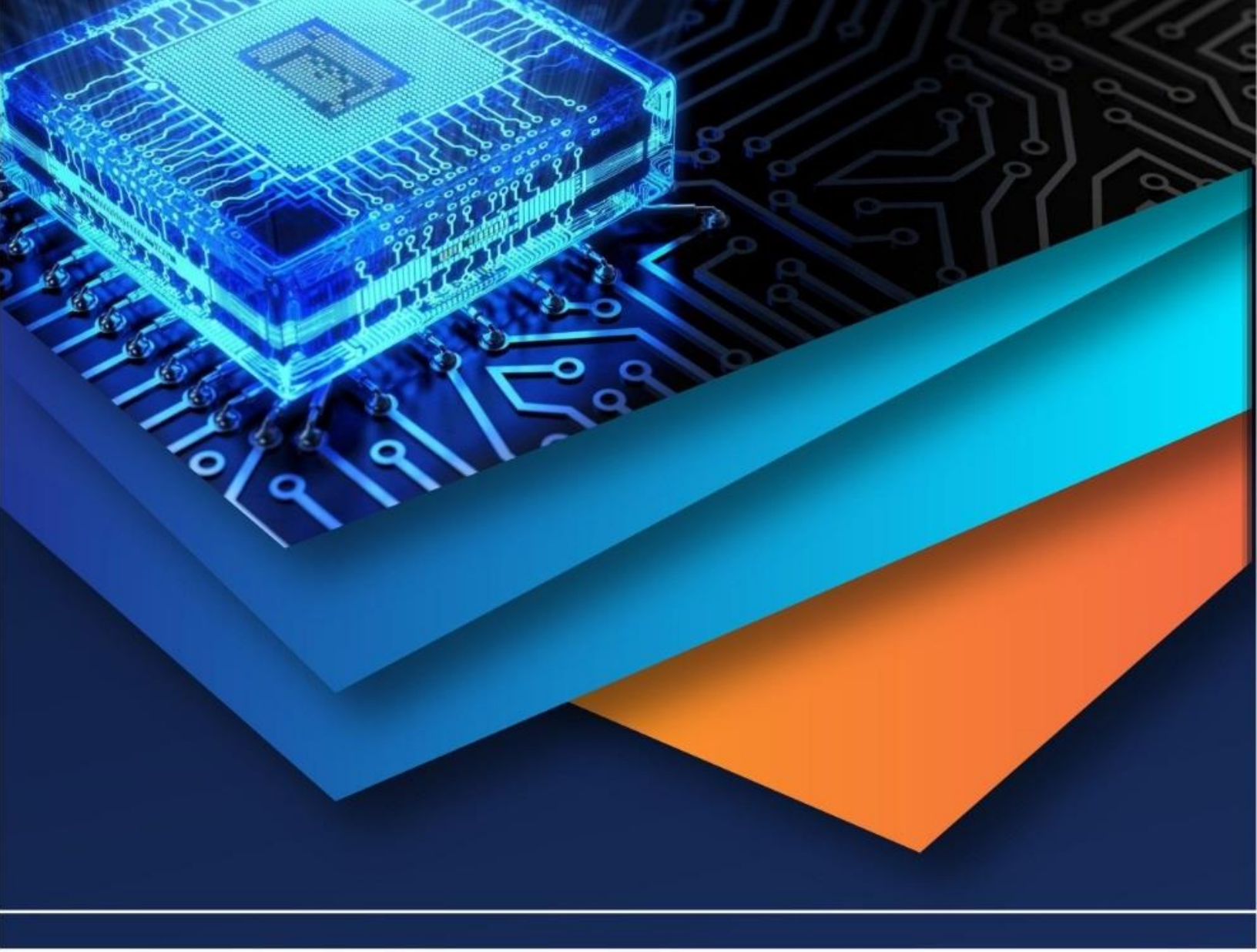
REFERENCES

[1] Petrini O (1991) Fungal endophytes of tree leaves. Microb Ecol Leaves 179:197.
[2] Petrini, O. 1991. Fungal endophytes of tree leaves. In: Andrews JH and Hirano SS, (Eds). Microbial leaf ecology. Spring Verlag. New York. 179-197.
[3] Petrini, O., 1991. Fungal endophytes of tree leaves. In: Fokkema, N.J., van den Heuvel 9 Eds.), Microbial; leaf ecology. Cambridge university press, Cambridge, UK, pp. 185-187.

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[4] Roza, L.V., Chanda, A., & Linz, J.E. 2011. Compartmentalisation and molecular trafficking in secondary metabolism: a new understanding of consolidation cellular processes. Fungal Genet. Biol. 48: 35–48.
[5] Faeth, S.H., and Fagan, W.F. 2002. Fungal Endophytes: Common Host Plant Symbiont but Uncommon Mutualism. Integ. and Comp. Biot. 43:360-368.
[6] Mousa, W.K., and Raizada, M.N. 2013. Review: The diversity of anti-microbial secondary metabolites produced by fungal endophytes: an interdisciplinary perspective. FCIMB. 4(65):1-18.
[7] Rodriguez, R. J., Arnold, J. F., Whiter Jr, A. E. and Redman, R.S., 2008. Review of Fungal endophytes: diversity and functional roles. New Phytol. 117.
[8] Shrivatsava, Tejas patel, 2007. Medicinal and aromatic plant science and biotechnology 1(1), 142-150.
[9] Selim, KA, El-Beih, A.A, AbdElRahman, TM, and ElDiwany, AI. 2012. Biology of endophytic fungi. Corr. Res. Environment. Apply. Mycol. 2(1):31–82.
[10] Zuber, A., Kowalczyk, M., Sekula, A., Mleczko, P., and Kupiec, T. 2011. The method used in species identification of hallucinogenic and other poisonous mushroom investigations.Prob. Foreign. Sci.86:151-161.
[11] Gandjar, I., dan R.A. Sjamsuridzal, W., dan Oetari, A. 2006. Mikologi dasar dan terapan, Yayasan Obor Indonesia, Jakarta.
[12] Gandjar, I., R.A. Samson, K. van den TweelVermeulen, Oetari, A dan Santoso, I. 1999. Pengenalan kapang tropik umum, Yayasan Obor Indonesia, Jakarta.
[13] Sedlar J., Sedlarova M., and Flusser J. 2009. Image Processing Methods for the Determination of Downy Mildew from optical Microscope Images. In: Kulpa K, Kaska W. (Eds) Signal Processing Symposium Proceedings, Warsaw University of Technology, Warsaw.
[14] Pitt, J.I. and A.D. Hocking, 2009. Fungi and Food Spoilage. 3rd Edn. Springer, USA. 519.
[15] Stone, J.K., Polishook J.D., and White J.F. 2004. Endophytic fungi. In: Biodiversity of Fungi. Inventory and Monitoring Methods. In Muller G.M., Bills G.F., Foster M.S.(Eds) Elsevier Academic Press, San Diego, United State of America. 241-270.

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