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Role of Fungal Endophytes in Plant Growth Promotion

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Abstract: ***Plant-associated microorganisms, such as fungal endophytes, arbuscular mycorrhizal fungi, and plant growth-promoting rhizobacteria, are well-documented for their function in encouraging crop productivity and extending stress forbearance. Fungal endophytes improve progress and healthiness of plants because of plant growth supporting type of rhizobacteria. They go through the plant tissues all the way across the root regions or aerial sections, through various parts as propagating radicles, secondary roots, stomata, or foliar way. Endophyte-plant-polymer decomposing enzymes for example cellulases, and pectinases perform a responsibility for their inner establishment and could be identified by immunological or in situ hybridization or cataloguing with reporter genes. Fungal endophytes act together biochemically as well as genetically with their native plant and produce osmolytes, osmo protectants, antioxidants, permitting the plants to alleviate the influences of different abiotic stress. Plant genes are controlled by endophyte, and the genes so extracted offer signs for the outcomes of endophytes in host plants. The present review describes bioprospecting of endophyte-plant interactions and discusses the way forward to recognise their molecular mechanisms and eventually the role of fungal endophytes. These are valuable representations to for learning about the genetic appearance of micro-organism inside the host plants, which are well-controlled and flexible. This supports in development of successful endophyte bioinoculants for abiotic stress as well as management of crop diseases. Fungal endophytes are general populations within the plant tissues and have been indicated to support plant development and wellbeing. Though, bit is understood regarding plant growth-promoting endophytes (PGPE). This review will help to take an account of role of fungal endophytes in plant grow promotion***

Keywords: Plant growth, fungal endophytes, abiotic and biotic stress,PGPE

I. INTRODUCTION

The methods regarding long term uses of organic and inorganic fertilizers alongside pesticides are instantly needed for the improvement of good crop production [1]. Remarkably, these applications always have adverse impact on quality of soil and promote to pollution in whole environment[2]. However just to reduce the harmful impacts of the traditional methods of agriculture, some of the advance approaches those cantered on microbial inoculation are just getting additional points of attention. Plants and microbes create a symbiotic association with advantages for both partners equally. More significantly, plant-microbe symbiosis impact on plant development and wellbeing which successfully enhances agricultural characteristics and enrich soil quality and nutrient cycling as well [3], [4], [5].

Usually, several microorganisms are found to obtain nutrients for their constant survival through collaboration with plants which may be independent, destructive (parasitism), or helpful (mutualism or symbiosis) to the host [6, 7]. Microbes that live within the plant tissues lacking doing substantial harm or obtaining compensation other than guaranteeing their position are believed as endophytes. Plants have been exalted by natural surroundings with differentiated population of endophytic microbes including useful bacteria, fungi, and actinomycetes.

Plant growth-promoting endophytes are responsible for inhabiting in plant tissues accelerates nutrients interchange and enzymatic activity [8], [9]. The dissemination of growth-promoting hormones generated by endophytic microbes to plant tissues completely encourages plant development [10]. Endophytes acquire essential capability to activate impenetrable phosphate and supply nitrogen to the host plants [11], [12]. Microbial endophytes colonize with the plant tissues without any symptomatic actions and accordingly they struggle with additional microscopic pathogens on the same biological recesses. They use their whole or part of the life cycle staying in the plant triggering no visual symptoms of disease

Hence, the recognized plant-endophyte togetherness enhances plant health by means of different processes presented by endophytes and possibly impacts the security of plant host versus microbial pathogens [13]. PGPE generate numerous bioactive composites along with numerous organic actions which could be defined as plant growth-promoting (PGP) agents directly or indirectly. Around most of the plants hold endophytes inside the tissues, though, existing data on PGPE and their natural actions is not comparable to

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the high-level dissemination of endophytes. A choice of understanding of the indigenous endophytes of plants might assist simplify their capabilities and possibility in improving plant development and determining a viable approach for crop production.
Endophytes have comprehensive role they play, are now deemed vital in the perception of their probable use to accomplish environmental advancements in the agro-food structure. Consequently, there is currently a technical ferment attempting to examine each facet of their contact with plants and concerned pathogens. Establishment of such valuable plant combined microorganisms is constantly getting consideration amongst technical people and at the understanding idea of productive industries due to their ability to improve plant condition and development

II. FUNGAL ENDOPHYTE

Fungi that live within living tissue lacking any symptoms of infection in their hosts are believed to be as fungal endophytes [14]. These are the most important participants of endophytic inhabitants that live completely within plant tissues and may be associated with all parts of plants. Each plant harbour at minimum one or additional endophytic fungi in the cosmos. In current years, they have been comprehensively explored in different geographical and climatical territories and were discovered to be abundant within plant internal tissues and have diversified species [15]. Scientists observed their essential functions in nutrients source, natural environment adaptation, protection of biotic and abiotic stresses, development promotion and improving community biodiversity of host plants [16]. They can also function as guardian in contrast to hunters and competitors of microbial pathogens. Earlier articles demonstrated that several grass species showed vegetative growth enhancement in the company of their fungal partners that have been principally attributed to expand fitness of host plants [17]. Nevertheless, later surveys have discovered that plant growth promotion may be recognized to the ejection of plant growth progressing secondary metabolites such as gibberellins, auxins, cytokinin secreted by the endophytic fungi in the rhizospheres. Literature analysis also explained that Plant Growth Promoting Fungi (PGPF) maintain plant growth through the invention of a several major enzymes such as ACCD, urease, catalase, etc., Previous details indicated that antibiotic resistant PGPE may be a useful resource of biocontrol agents [18].

III. TAXONOMY OF FUNGAL ENDOPHYTES

Fungal endophytes might be categorized mostly into environmental and diversity categories or based on functional roles. They have been arranged into two major groups as

1) Clavicipitaceous fungal endophytes are usually popular in grasses
2) non-clavicipitaceous are leading with vascular and non-vascular plants [19].

Fungal endophytes are further classified based on numerous standards such as: On the basis of the host plant part affected, source of nutrition, mode of transmission, expression of infection, mode of reproduction [20].

A. On The Basis Of The Host Plant Part Affected

Fungal endophytes can be classified as root and foliar endophytes based on part of plant. For example, in a survey revealed that *Pochonia chlamydosporia* and *B. bassiana* were identified within the leaves and stem, though *Metarhizium* spp. was observed in roots. [21]. Fungal endophytes that infect roots as *Piriformospora indica*, *Metarhizium* spp., *Fusarium* spp., and *Glomus* spp., are example of root endophytes [22]. Others that enter leaves and stems are identified as foliar endophytes [23].
Fungal endophytes are normally recognised based on their structural features. However, apart from structural classification, there are recommendations that, isolation and molecular classification of fungal endophytes are incredibly significant for their identification [24]' that can be conducted out through with the amplification and sequencing a little portion of fungal DNA [25].

B. On the basis of source of nutrition,

Based on this criteria, fungal endophytes can be categorized as necrotrophs, or as biotrophs. Necrotrophs, confound the cells of host to grow on the dead tissues while biotrophs will develop and acquire nutrients living host tissues. (26). The fungi-plant interfaces happen when fungal endophytes get carbon supply from hosts, and in replacement for the energy supplies, obtained from the hosts, deliver advantages to the plant (27). Though, due to evolutionary and environmental variations, probably some fungal endophytes shifting from biotrophic to necrotrophic lifestyle [26]. For example, *Leptosphaeria maculans* following asymptomatic in healthful *Arabidopsis thaliana* plants turn out to be a necrotrophic pathogen after the host was stressed [28].

C. On the Basis of Mode of Transmission

Fungal endophytes might be classified **as**

1) Vertically–transmitted, those are moved immediately from the host (parents) to their offspring [29. Actual endophytes as *Neotyphodium* are mainly have vertical transmission over seeds [30]. When mode of transmission is vertical due to seeds, they are defined to as seed-transmitted endophytes [31].
2) Horizontally transmitted endophytes are transported among various entities in a particular inhabitant. This mode of transmission is common with fungal endophytes that contaminate plants through airborne spores (30). Endophytes are normally increased through vegetative parts, or transmission by spores, known as spore-transmitted endophytes [32].

D. On the Basis of Expression of Infection,

Endophytes are categorised with symptoms (symptomatic) and without symptoms (asymptomatic) [33] Several of fungal endophytes contaminate aerial internal plant tissues deprived of presenting symptoms. Countless consideration is engrossed on these endophytes as they are general and have enormous variety and numerous functions [34]. For example, *Fusarium* spp. was recognized as asymptomatic as it was established to direct without symptoms in banana cultivar's cord roots [35]. Though in exceptional instances, symptomatic endophytes can be classified as asymptomatic as the host plant is resilient to the fungi. Though, as previously acknowledged, a variation in ecological situations might trigger an unexpected change in the performance of fungal endophytes without symptoms. A perfect case is the example of fungi that were obtained as in form of endophytes having no symptoms , however, developed infective in altered ecological situations [26]. Hence the stage of the host carrying the fungus and the ecological situations possess a bigger part to participate in establishing whether a fungal endophyte functions as a symptomless endophyte or else like a symptom-creating type of plant pathogen [36].

E. On the Basis of Mode of Reproduction

Fungal endophytes are categorized **as** **a**sexual and sexual [37]. The *Epichloë* endophytes have been separated in *Epichloë* and *Neotyphodium*, that are having mode of reproduction as sexual and asexual correspondingly [38].

IV. FUNGAL ENDOPHYTES: BIOCONTROL, BIOFERTILIZATION AND STRESS TOLERANCE

Most of regions on the globe have suffered a decline in water amount and an increase in intensity of soil salinity and draught along with other troubles linked to the disproportionate uses of soil, deforestation, and improper irrigation methods [39]. A technique of solving these types of concerns is the growth of diverse types of plant by producing wild plants. Still, the genomic processes participating in stress forbearance are inadequately understood, and an important part is not believed in this procedure: the symbiotic relationship of plants and microbes [40]. The fungal endophytes that survive in wild plants can be strictly adapted, might be missing through taming; consequently, fungi are affected by mislaying their safe position and plants are lacking a relationship that might recover their capability to overcome ecological encounters (39).

For endophytes, the innermost part of the plant is a secured position that includes the essential nutrients for existence and development of fungi in addition to exhibiting lower competition with other microbes. Thus, in swap for this secure position, fungi enhance fitness of plant by various methods [41].

The advantages of plant establishment by fungal endophytes be able to happen in direct and/or indirect manner, and the variation amongst them is complicated [42]. Amongst the direct process of growth promotion, the highly valuables are the asset of nutrients and the creation of phytohormones, though forbearance to stresses (biotic and abiotic), together with fighting in contradiction of pathogens, is measured as an indirect facet in the promotion of growth [43].The use of fungal endophytes is really beginning, and in the next segments, the direct and indirect processes by which these microbes can aid in plant health will be detailed, including a perspective of their use in agricultural processes soon.

Current studies revealed two or even additional capabilities offered by the similar strains instead of only a one ability, as the rise in nutrient uptake or phytohormone production [44]. The strains were established in leguminous plants and were proved to encourage development-improving criteria as dry matter (shoot and roots) and the accessibility of vital nutrients such as Phosphorous and Nitrogen

The direct process of contact with fungal endophytes will include asset of nutrients and in the quantity of phytohormones in the plant, which is completely associated with expansion of biomass creation, increase of root structure progress, mass generation plant elevation, and produce. As of these advantages, they can be described in form of biofertilizers [45].

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A. Acquisition of Nutrients

Fungal endophytes are capable to enhance the uptake of macronutrients, for example magnesium, potassium, nitrogen and phosphorus, or micronutrients, such as copper, iron and zinc, from the soil as well as from organic matter and rise the source of these nutrients to the host Plant [46].In context to the processes of nutrient uptake movement , yet no exact metabolic pathways and molecules involved in many processes are described

B. Production of phytohormones

Fungal endophytes are competent to generate different phytohormones such as cytokinin, gibberellins (GAs) and auxins. The capability of phytohormone making by these endophytes is not explored clearly, particularly for gibberellins, although these particles are as significant as chemical indicating and heralds for plant development in various ecological situations [47]. The primary auxin made by fungi is indole-3-acetic acid. Auxins are the most important regulators of plant development and produce numerous helpful impacts on shoot and root growth, such as the reactions of tropism, cell division and cell elongation, differentiation of vascular tissue and beginning of the root development procedure [48] Gibberellins are important in various plant reactions, as stem elongation, sexual expression, seed germination, senescence fruit formation and flourishing, (49).

C. Indirect Process of contact with Fungal Endophytes

1) *Activation of Systemic Resistance:* Fungal endophytes can assist hosts in developing their immunity, thus encouraging the stimulation of induced systemic resistance pathways, which might coincide with that of acquired systemic resistance as both can enhance plant development together [50] and protect against pests and pathogens .
2) *Production of Antibiotics and Secondary Metabolites:* In addition to encouraging the making of molecules of immunity by the plant itself, fungal endophytes are a significant reservoir of particles favouring to host. These are outstanding creators of composites including alkaloids, steroids, terpenoids, peptides, polyketones, flavonoids, quinols, phenols, chlorinated compounds, and volatile organic compounds (VOCs), having action versus pathogens and herbivores, [51]. Furthermore, survey recommend the production of composites along with antiviral, antibacterial, antifungal and insect activity [52].
3) *Protection Against Biotic and Abiotic Stresses:* Ecological deprivation by farming practices and worldwide environment transformation uncovers plants to increasingly tricky situations for their development and conservation as well. Furthermore, the condition is much added complicated for crops as greater harvests are progressively needed. In this situation, it is obvious that certain assistance is needed for excellent quality plant growth, and fungal endophytes are a hopeful option for plant safeguard against biotic and abiotic stresses. Fungal endophytes are capable to battle with abiotic stresses, like drought, hot and cold atmosphere, saltiness, and poisonous heavy metals [47]. For biotic stress safety, fungal endophytes are accountable for the stimulation of ISR and ASR, which generate metabolites for pathogens; likewise, parasitism or competition can happen to prevent infection and herbivory [41].The benefits of consuming fungi and other microbes in farming are previously well recognised and comprise of better biosafety, fewer ecological and social health hazard, specificity with the mark, effectiveness even in small quantities, multiplication, no resistance, no advancement of the choice of resistant pests, usage in IPM or in the old-style farming scheme [42].The fungi as biological control, particularly against entomopathogens, has developed well-known as natural substitutes to chemicals. The restriction will be mostly to UV radiation and minimal humidity observed in the cultivated atmosphere, adding to difficulties associated with use in the area [53]. A further recent idea is that disease alteration by endophytes is perspective-related, as it depends on biotic and abiotic factors of the atmosphere, host /or pathogens. Differences in physical parameters can impact the aggressive action of endophytes. At the same time, the plant's personal microflora act together with the endophyte and might be accountable for differences in reaction towards pathogen [50]. Worldwide, data on the application of organic products has become more helpful and with an improved knowledge of what they comprise of and the improvements of their usage, adding to the commercial feasibility of their purpose, their usage has increased.

V. CONCLUSION

The plant development encouraging capability of fungal endophytes might be appropriate to their ability to secret improved quantities of several beneficial development boosting metabolites and thus help their hosts to stay alive under stress (biotic and abiotic) condition. Several findings of previous studies motivate to spread the knowledge on the selected fungal endophytes to grow a sturdy Bioagent with large pertinence to multi field and henceforth developing as a flourishing bio inoculum proceeding towards organic food crops for a healthier future by dropping the life-threatening uses of chemicals.

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With the increasing apprehension nearby the justification of ecological influences triggered by farming procedures in countryside and in the exploration for improved nourishments, chemical free composites injurious to well-being, research is evolving concerning the use of microbes for this purpose. The capability of endophytic establishment of crops determined by numerous fungi, with those traditionally employed for pest management, has demonstrated to be an extremely probable system to achieve the preferred sustainability in farming sector. As indicated in this review article, fungal endophytes are useful as they make available numerous direct and indirect advantages for crops, and it is feasible to suppose that no artificial particle is capable to give such a fantastic variety of helpful relations as these microbes do. Therefore, the usage of fungal endophytes validates to be an option of excellent ability in the areas of biocontrol, bio stimulation and biofertilization, explaining that these microbes are a strong means for exploration and operations and play an incredibly significant role in growth promotion of crops.

REFERENCES

[1] Bokhtiar S.M., Sakurai K. Effect of application of inorganic and organic fertilizers on growth, yield, and quality of sugarcane. *Sugar Tech.* 2005;7(1):33–37. [Google Scholar]
[2] Aktar M.W., Sengupta D., Chowdhury A. Impact of pesticides use in agriculture: their benefits and hazards. *Interdiscip Toxicol.* 2009;2(1):1–12. [PMC free article] [PubMed] [Google Scholar]
[3] Khan A.L., Waqas M., Khan A.R., Hussain J., Kang S.-M., Gilani S.A. Fungal endophyte Penicillium janthinellum LK5 improves growth of ABA-deficient tomato under salinity. *World J Microbiol Biotechnol.* 2013;29(11):2133–2144. [PubMed] [Google Scholar]
[4] Karthik C., Oves M., Thangabalu R., Sharma R., Santhosh S.B., Indra Arulselvi P. Cellulosimicrobium funkei-like enhances the growth of *Phaseolus vulgaris* by modulating oxidative damage under Chromium (VI) toxicity. *J Adv Res.* 2016;7(6):839–850. [PMC free article] [PubMed] [Google Scholar]
[5] Puri A., Padda K.P., Chanway C.P. Seedling growth promotion and nitrogen fixation by a bacterial endophyte *Paenibacillus polymyxa* P2b–2R and its GFP derivative in corn in a long-term trial. *Symbiosis.* 2016;69(2):123–129. [Google Scholar]
[6] Shen, H. W. Ye, L. Hong et al., "Progress in parasitic plant biology: Host selection and nutrient transfer," *The Journal of Plant Biology*, vol. 8, no. 2, pp. 175–185, 2006.View at: Publisher Site | Google Scholar
[7] Thrall P. H., Hochberg, M. E. Burdon, J. J. and Bever, J. D. "Coevolution of symbiotic mutualists and parasites in a community context," *Trends in Ecology & Evolution*, vol. 22, no. 3, pp. 120–126, 2007.View at: Publisher Site | Google Scholar
[8] Khan A.R., Ullah I., Waqas M., Shahzad R., Hong S.-J., Park G.-S. Plant growth-promoting potential of endophytic fungi isolated from Solanum nigrum leaves. *World J Microbiol Biotechnol.* 2015;31(9):1461–1466. [PubMed] [Google Scholar]
[9] Murphy B.R., Doohan F.M., Hodkinson T.R. Yield increase induced by the fungal root endophyte *Piriformospora indica* in barley grown at low temperature is nutrient limited. *Symbiosis.* 2014;62(1):29–39. [Google Scholar]
[10] Lin L., Xu X. Indole-3-acetic acid production by endophytic streptomyces sp. En-1 isolated from medicinal plants. *Curr Microbiol.* 2013;67(2):209–217. [PubMed] [Google Scholar]
[11] Matsuoka H., Akiyama M., Kobayashi K., Yamaji K. Fe and P solubilization under limiting conditions by bacteria isolated from *Carex kobomugi* Roots at the Hasaki Coast. *Curr Microbiol.* 2013;66(3):314–321. [PubMed] [Google Scholar]
[12] Shi Y., Lou K., Li C. Growth promotion effects of the endophyte *Acinetobacter johnsonii* strain 3–1 on sugar beet. *Symbiosis.* 2011;54(3):159–166. [Google Scholar]
[13] Malhadas C., Malheiro R., Pereira J.A., de Pinho P.G., Baptista P. Antimicrobial activity of endophytic fungi from olive tree leaves. *World J Microbiol Biotechnol.* 2017;33(3):46. [PubMed] [Google Scholar]
[14] Rodriguez R. J., J. F. Jr., A. E. Arnold, and R. S. Redman, "Fungal endophytes: diversity and functional roles," *New Phytologist*, vol. 182, no. 2, pp. 314–330, 2009.View at: Publisher Site | Google Scholar
[15] A. Tanwar and A. Aggarwal, "Multifaceted potential of bioinoculants on red bell pepper (F1 hybrid, Indam Mamatha) production," *Journal of Plant Interactions*, vol. 9, no. 1, pp. 82–91, 2014.View at: Publisher Site | Google Scholar
[16] H.-Y. Li, D.-W. Li, C.-M. He, Z.-P. Zhou, T. Mei, and H.-M. Xu, "Diversity and heavy metal tolerance of endophytic fungi from six dominant plant species in a Pb-Zn mine wasteland in China," *Fungal Ecology*, vol. 5, no. 3, pp. 309–315, 2012.View at: Publisher Site | Google Scholar
[17] B. R. Glick, "Bacteria with ACC deaminase can promote plant growth and help to feed the world," *Microbiological Research*, vol. 169, no. 1, pp. 30–39, 2014.View at: Publisher Site | Google Scholar
[18] F. Yasmin, R. Othman, K. Sijam, and M. S. Saad, "Characterization of beneficial properties of plant growth-promoting rhizobacteria isolated from sweet potato rhizosphere," *African Journal of Microbiology Research*, vol. 3, no. 11, pp. 815–821, 2009.View at: Google Scholar
[19] Rodriguez RJ, White Jr JF, Arnold AE, Redman RS. 2009. Fungal endophytes: diversity and functional roles. New Phytologist. 182(2):314–330. doi:https://doi.org/10.1111/j.1469-8137.2009.02773.x. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]
[20] Purahong, W., and Hyde, K. D. (2011). Effects of fungal endophytes on grass and non-grass litter decomposition rates. *Fungal Divers.* 47, 1–7. doi: 10.1007/s13225-010-0083-8 CrossRef Full Text | Google Scholar
[21] Behie, S. W., and Bidochka, M. J. (2014). Ubiquity of insect-derived nitrogen transfer to plants by endophytic insect-pathogenic fungi: an additional branch of the soil nitrogen cycle. *Appl. Environ. Microbiol.* 80, 1553–1560. doi: 10.1128/AEM.03338-13 PubMed Abstract | CrossRef Full Text | Google Scholar
[22] Wyrebek, M., Huber, C., Sasan, R. K., and Bidochka, M. J. (2011). Three sympatrically occurring species of *Metarhizium* show plant rhizosphere specificity. *Microbiology* 157, 2904–2911. doi: 10.1099/mic.0.051102-0 PubMed Abstract | CrossRef Full Text | Google Scholar
[23] Meyling, N. V., Thorup-Kristensen, K., and Eilenberg, J. (2011). Below-and aboveground abundance and distribution of fungal entomopathogens in experimental conventional and organic cropping systems. *Biol. Control* 59, 180–186. doi: 10.1016/j.biocontrol.2011.07.017 CrossRef Full Text | Google Scholar

[24] Lu, L., Cheng, B., Du, D., Hu, X., Peng, A., Pu, Z., et al. (2015). Morphological, molecular and virulence characterization of three *Lencanicillium* species infecting Asian citrus psyllids in Huangyan citrus groves. *J. Invertebr. Pathol.* 125, 45–55. doi: 10.1016/j.jip.2015.01.002 PubMed Abstract | CrossRef Full Text | Google Scholar

[25] Chen, L., Li, X., Li, C., Swoboda, G. A., Young, C. A., Sugawara, K., et al. (2015). Two distinct *Epichloë* species symbiotic with *Achnatherum inebrians*, drunken horse grass. *Mycologia* 107, 863–873. doi: 10.3852/15-019 PubMed Abstract | CrossRef Full Text | Google Scholar

[26] Delaye, L., García-Guzmán, G., and Heil, M. (2013). Endophytes versus biotrophic and necrotrophic pathogens—are fungal lifestyles evolutionarily stable traits? *Fungal Divers.* 60, 125–135. doi: 10.1007/s13225-013-0240-y CrossRef Full Text | Google Scholar

[27] Behie, S. W., and Bidochka, M. J. (2014). Ubiquity of insect-derived nitrogen transfer to plants by endophytic insect-pathogenic fungi: an additional branch of the soil nitrogen cycle. *Appl. Environ. Microbiol.* 80, 1553–1560. doi: 10.1128/AEM.03338-13 PubMed Abstract | CrossRef Full Text | Google Scholar

[28] Junker, C., Draeger, S., and Schulz, B. (2012). A fine line – endophytes or pathogens in *Arabidopsis thaliana*. *Fungal Ecol.* 5, 657–662. doi: 10.1016/j.funeco.2012.05.002 CrossRef Full Text | Google Scholar

[29] Saikkonen, K., Ion, D., and Gyllenberg, M. (2002). The persistence of vertically transmitted fungi in grass metapopulations. *Proc. Biol. Sci.* 269, 1397–1403. doi: 10.1098/rspb.2002.2006 PubMed Abstract | CrossRef Full Text | Google Scholar

[30] Hartley, S. E., and Gange, A. C. (2009). Impacts of plant symbiotic fungi on insect herbivores: mutualism in a multitrophic context. *Annu. Rev. Entomol.* 54, 323–342. doi: 10.1146/annurev.ento.54.110807.090614 PubMed Abstract | CrossRef Full Text | Google Scholar

[31] Bennett, R., Hutmacher, R., Davis, R., and Bennett, R. (2008). Seed transmission of *Fusarium oxysporum* f. sp. *vasinfectum* race 4 in California. *J. Cotton Sci.* 12, 160–164. Google Scholar

[32] Faeth, S. H., and Fagan, W. F. (2002). Fungal endophytes: common host plant symbionts but uncommon mutualists. *Integr. Comp. Biol.* 42, 360–368. doi: 10.1093/icb/42.2.360 PubMed Abstract | CrossRef Full Text | Google Scholar

[33] Pinto, L. S. R. C., Azevedo, J. L., Pereira, J. O., Vieira, M. L. C., and Labate, C. A. (2000). Symptomless infection of banana and maize by endophytic fungi impairs photosynthetic efficiency. *New Phytol.* 147, 609–615. doi: 10.1046/j.1469-8137.2000. 00722.x CrossRef Full Text | Google Scholar

[34] Arnold, A. E., and Lutzoni, F. (2007). Diversity and host range of foliar fungal endophytes: are tropical leaves biodiversity hotspots? *Ecology* 88, 541–549. PubMed Abstract | Google Scholar

[35] Sikora, R. A., Pocasangre, L., Zum Felde, A., Niere, B., Vu, T. T., and Dababat, A. (2008). Mutualistic endophytic fungi and in-planta suppressiveness to plant parasitic nematodes. *Biol. Control* 46, 15–23. doi: 10.1016/j.biocontrol.2008.02.011 CrossRef Full Text | Google Scholar

[36] Porras-Alfaro, A., and Bayman, P. (2011). Hidden fungi, emergent properties: endophytes and microbiomes. *Annu. Rev. Phytopathol.* 49, 291–315. doi: 10.1146/annurev-phyto-080508-081831 PubMed Abstract | CrossRef Full Text | Google Scholar

[37] Brem, D., and Leuchtmann, A. (2001). Epichloë grass endophytes increase herbivore resistance in the woodland grass *Brachypodium sylvaticum*. *Oecologia* 126, 522–530. doi: 10.1007/s004420000551 PubMed Abstract | CrossRef Full Text | Google Scholar

[38] Schardl, C., and Craven, K. (2003). Interspecific hybridization in plant-associated fungi and oomycetes: a review. *Mol. Ecol.* 12, 2861–2873. doi: 10.1046/j.1365-294X.2003. 01965.x PubMed Abstract | CrossRef Full Text | Google Scholar

[39] Lugtenberg BJJ, Caradus JR, Johnson LJ. 2016. Fungal endophytes for sustainable crop production. FEMS Microbiol Ecol. Dec; 92(12):17. doi: https://doi.org/10.1093/femsec/fiw194. [Crossref], [Web of Science ®], [Google Scholar]

[40] Chadha N, Mishra M, Rajpal K, Bajaj R, Choudhary DK, Varma A. 2015. An ecological role of fungal endophytes to ameliorate plants under biotic stress. Arch Microbiol. Sep; 197(7):869–881. doi:https://doi.org/10.1007/s00203-015-1130-3. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

[41] Chitnis VR, Suryanarayanan TS, Nataraja KN, Prasad SR, Oelmuller R, Shaanker RU. 2020. Fungal endophyte-mediated crop improvement: the way ahead. Front Plant Sci. Oct; 11:10. doi:https://doi.org/10.3389/fpls.2020.561007. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

[42] Berg G. 2009. Plant-microbe interactions promoting plant growth and health: perspectives for controlled use of microorganisms in agriculture. Appl Microbiol Biotechnol. Aug; 84(1):11–18. doi:https://doi.org/10.1007/s00253-009-2092-7. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

[43] Souza BD, dos Santos TT. 2017. Endophytic fungi in economically important plants: ecological aspects, diversity, and potential biotechnological applications. Journal of Bioenergy and Food Science. Apr-Jun; 4(2):113–126. doi: https://doi.org/10.18067/jbfs.v4i2.121. [Crossref], [Web of Science ®], [Google Scholar]

[44] Bader AN, Salerno GL, Covacevich F, Consolo VF. 2020. Native trichoderma harzianum strains from Argentina produce indole-3 acetic acid and phosphorus solubilization, promote growth and control wilt disease on tomato (solanum lycopersicum L.). Journal of King Saud University - Science. Jan; 32(1):867–873. doi:https://doi.org/10.1016/j.jksus.2019.04.002. [Crossref], [Web of Science ®], [Google Scholar]

[45] Bamisile BS, Dash CK, Akutse KS, Keppanan R, Wang LD. 2018. Fungal endophytes: beyond herbivore management. Front Microbiol. Mar; 9:11. doi:https://doi.org/10.3389/fmicb.2018.00544. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

[46] Rana KL, Kour D, Kaur T, Devi R, Yadav AN, Yadav N, Dhaliwal HS, Saxena AK. 2020. Endophytic microbes: biodiversity, plant growth-promoting mechanisms and potential applications for agricultural sustainability. Antonie Van Leeuwenhoek International Journal of General and Molecular Microbiology. Aug; 113(8):1075–1107. doi:https://doi.org/10.1007/s10482-020-01429-y. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

[47] Khan AL, Hamayun M, Kang SM, Kim YH, Jung HY, Lee JH, Lee IJ. 2012. Endophytic fungal association via gibberellins and indole acetic acid can improve plant growth under abiotic stress: an example of paecilomyces formosus LHL10. BMC Microbiol. Jan; 12(1):14. doi:https://doi.org/10.1186/1471-2180-12-3. [Crossref], [PubMed], [Google Scholar]

[48] Jaroszuk-Ściseł J, Kurek E, Trytek M. 2014. Efficiency of indoleacetic acid, gibberellic acid and ethylene synthesized in vitro by fusarium culmorum strains with different effects on cereal growth. Biologia. 69(3):281–292. doi: https://doi.org/10.2478/s11756-013-0328-6. [Crossref], [Web of Science ®], [Google Scholar]

[49] Bömke C, Tudzynski B. 2009. Diversity, regulation, and evolution of the gibberellin biosynthetic pathway in fungi compared to plants and bacteria. Phytochemistry. 70(15–16):1876–1893. doi:https://doi.org/10.1016/j.phytochem.2009.05.020. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

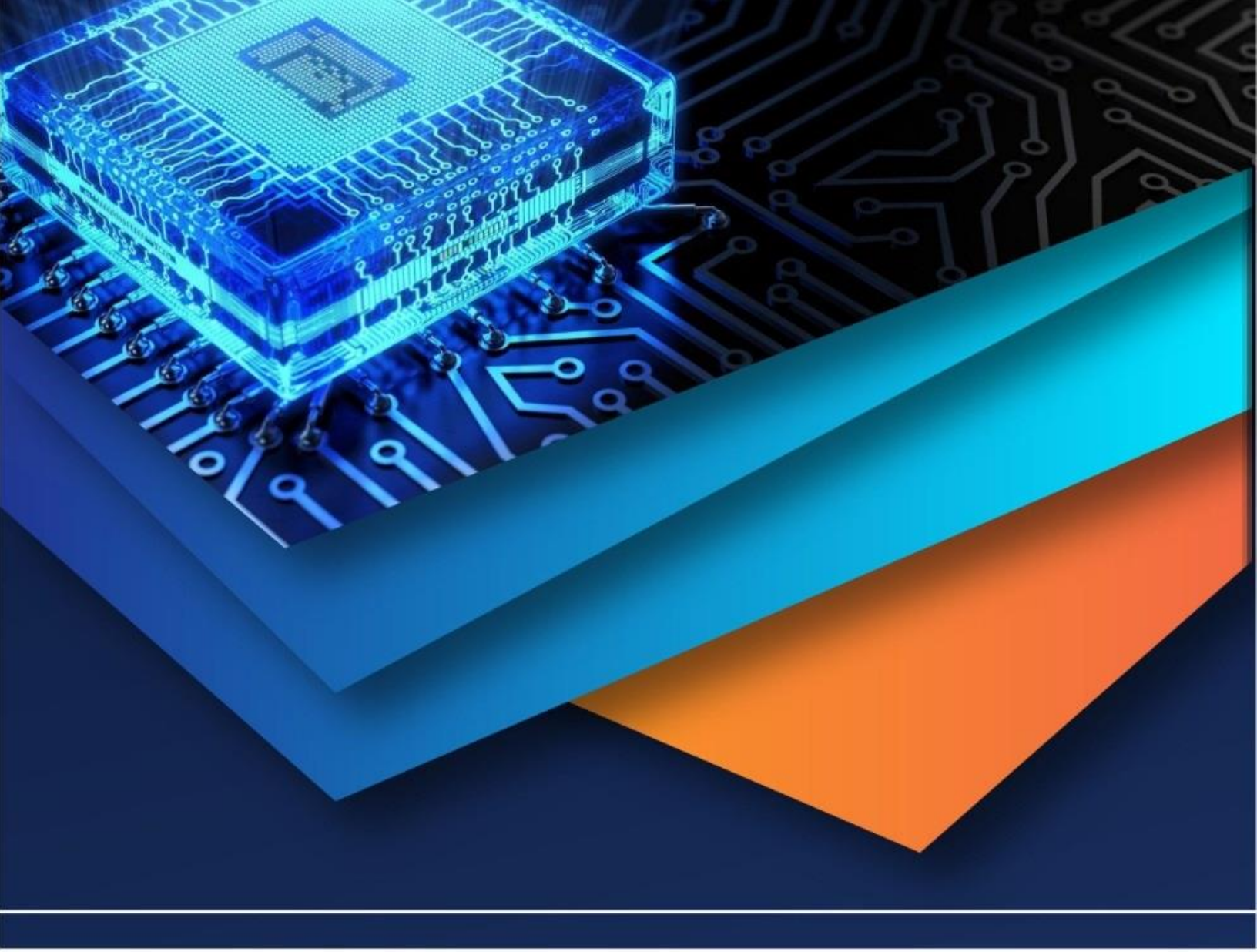
[50] Busby PE, Ridout M, Newcombe G. 2016. Fungal endophytes: modifiers of plant disease. Plant Mol Biol. Apr; 90(6):645–655. doi:https://doi.org/10.1007/s11103-015-0412-0. [Crossref], [PubMed], [Web of Science ®], [Google Scholar]

[51] Kaddes A, Fauconnier ML, Sassi K, Nasraoui B, Jijakli MH. 2019. Endophytic fungal volatile compounds as solution for sustainable agriculture. Molecules. Mar; 24(6):16. doi: https://doi.org/10.3390/molecules24061065. [Crossref], [Web of Science ®], [Google Scholar]

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[52] Latz MAC, Jensen B, Collinge DB, Jorgensen HJL. 2018. Endophytic fungi as biocontrol agents: elucidating mechanisms in disease suppression. Plant Ecol Divers. Nov; 11(5–6):555–567. doi: https://doi.org/10.1080/17550874.2018.1534146. [Taylor & Francis Online], [Web of Science ®], [Google Scholar]

[53] Vega V. 2018. The use of fungal entomopathogens as endophytes in biological control: a review. Mycologia. 110(1):4–30. doi: https://doi.org/10.1080/00275514.2017.1418578. [Taylor & Francis Online], [Web of Science ®], [Google Scholar]



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