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Taxonomy and Distribution of Recent Benthic Foraminifera from the Savitri Estuary, West Coast of India, Maharashtra

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Abstract: Foraminifera have thrived in a wide range of aquatic habitats, including the depths of the oceans, brackish lagoons, estuaries, and, on rare occasions, in freshwater streams and lakes. In the present study, environmental variables and benthic foraminiferal assemblages were studied using 17 samples obtained from various sites along the Savitri River. These samples comprised live specimens, as well as surface and subsurface (shallow core) samples. To know their distribution in, the present study has been undertaken nine stations starting from Velas to Umroli. Benthic foraminiferal taxa belonging to six sub orders (Textularina, Milolina, Lagenina and Rotalina), 16 super families, 20 families, 25 genera and 31 species have been identified. Among the 31 species, five species *Lagena semistriata* Williamson, *Bolivina striatula* d' Orbigny, *Rosalina bradyi* Cushman, *Ammonia tepida* and *Elphidium advenum* Cushman are considered to be abundant species of the present study. Higher diversity and population of foraminifera is noticed in the samples collected in the Bankot Jetty, because of their favourable niche. As moving towards landward locations, the lower diversity and population are observed. The substrate of present study area is seen to consist of Silty clay, Sand, Silty sand and Sand. Among these, silty sand provides the most favourable conditions for a higher population density. The primary ecological factors influencing the distribution of foraminifera in this area are the organic matter content and the characteristics of the substrate.

Keywords: Benthic foraminifera, Savitri Estuary, West Coast of India

I. INTRODUCTION

Benthic foraminifera serve as a valuable resource for comprehending marine environmental conditions due to their accessibility and their predominant presence in the uppermost layers of marine and estuarine sediments (Schafer et al., 1991). Furthermore, they provide a wealth of information regarding the ecology of diverse marine habitats. Consequently, they are recognized as effective indicators of ecological conditions (Badawi and El-Menhawey, 2016). Foraminifera have been effective inhabitants of all aquatic environments, including deep oceans, brackish water lagoons, estuaries, and occasionally freshwater streams and lakes. Due to their well-documented high fossilization potential, foraminifera tend to be abundant in marine sediments, making the collection and separation of these organisms from sediment samples relatively straightforward (Murray, 1991).

Numerous research efforts have employed the abundance and distribution of benthic foraminifera as indicators to investigate environmental factors such as productivity, dissolved oxygen levels, the thermohaline characteristics of the water column, and the circulation of bottom water (Badawi and El- Menhawey, 2016; Debenay et al., 2005; Duplessy et al., 1988; Mackensen et al., 1993; Hemleben and Bijma, 1994; Schmiedl and Leuschner, 2005; Bickert and Wefer, 1999; Berkeley et al., 2007). They constitute as much as 80% of the total biomass of protists and play a crucial role in the food web (Li et al., 2014; Nomaki et al., 2008; Wukovits et al., 2018). Foraminifera represent an important biological element of sandy beach ecosystems, with their shell content in certain regions potentially reaching as high as 95% of the sand composition (Barbieri et al., 2006; Dong et al., 2020). They serve as inherent indicators for quantitatively assessing the pollution levels in the marine environment caused by residential and industrial waste (Choi and An, 2012; Abu-Zied et al., 2013; Dimiza et al., 2022). The characteristics of their environmental dependency contribute to the distinction between recent and paleoenvironments (Kemp et al., 2020; Minhat et al., 2021). Consequently, it is crucial to define and comprehend the regional distribution of foraminifera.

A. Study Area

The Savitri River, located in the northernmost part of the Raigad district, serves as a natural boundary for approximately 24 miles between Raigad and Kolaba districts.

The estuarine area of this district is characterized by relatively fertile soils, which contribute to its status as the most agriculturally productive land in the region. The Savitri River runs in a northwest-southeast direction, indicating a structural influence in the tidal areas of the waterways. The entrance to the Savitri Estuary is characterized by bluffs that extend into the sea on both sides. Moving upstream from the river's mouth, the landscape becomes significantly more appealing, as the surrounding hills create a scenic view. As one travels further inland, the elevated table-land recedes from the riverbanks, giving way to expansive alluvial lowlands interspersed with patches of mangrove swamps. However, as one approaches Mahad, the riverbanks become flat and less visually stimulating.

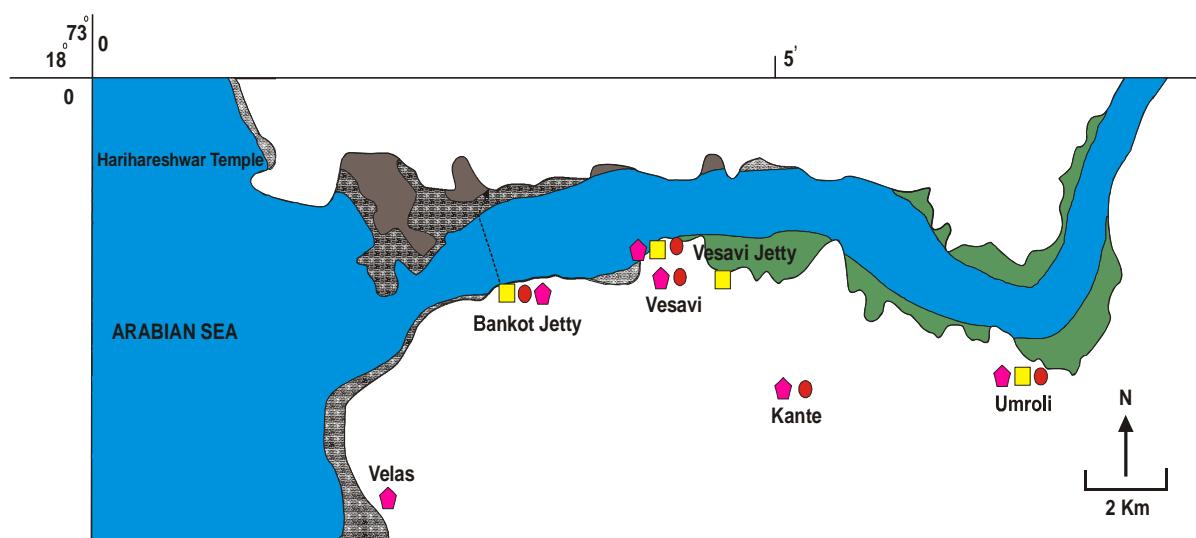
The Coastal Division of Konkan classifies the Savitri River as part of the Mesotidal Coast, characterized by a Spring tide range of 3.5 to 2 meters and a Neap tide range of 2 to 1.5 meters, with a tidal incursion limit of 25 kilometers in its tidal rivers (Karlekar, 1981). The primary sedimentary environments found in the creeks and estuaries of Konkan include marsh and swamp edges, high and low tide flats, sand lenses, sand banks, islands, and scoured channels (Karlekar et al., 1993). The sedimentation processes within these estuaries are influenced by various factors, including the length of the tidal inlet, the tidal range, and the flocculation process.

Table 1 - Sampling: Savitri Estuary (Southern bank), Ratnagiri District

Sr. No.	Locality	Location	Surface/ Subsurface	Lithology	Vegetation	Species Name	Family	Remarks
1)	Umroli	18° 08' 53" N 73° 17' 31" E	Live Surface Short core	Clay	Mangroves	<i>Aegiceras corniculata</i> (L.) Blanco	Myrsinaceae	Samples are collected about 10 km upstream from Savitri mouth
2)	Kante	18° 13' 07" N 73° 05' 42" E	Surface Short core	Clay	Two different types of Mangroves	<i>Avicennia marina</i> (Forssk.) Denkschr. <i>Sonneratia apetala</i> Buch.-Ham.	Avecenniaceae Sonneratiaceae	Landward end of Mangroves
3)	Veshavi Jetty	18° 14' 20" N 73° 15' 21" E	Live Surface Short core	Clay	Mangroves	—	—	Landward end spot is located about 5 ft. from lowest stand of MSL along the Jetty.
4)	Bankot Jetty	18° 13' 02" N 73° 04' 34" E	Live Surface Short core	Sandy Clay	Mangroves are Scanty	—	—	Sand content goes on increasing towards the mouth of the river

5)	Velas	18° 12' 43" N 73° 15' 32" E	Surface	Clay/Sand	Very Small patches of Mangroves	Lumnitzera racemosa Willd.	Combretaceae	Plantation is done on the Stabilized dune
6)	Vesavi (Along the road cutting)	17° 59' 34" N 73° 18' 41" E	Live Surface Short core	Clay	Thick Mangroves	Sonneratia alba J.Sm.	Sonneratiaceae	It is indicative of Landward end of Mangroves

Savitri Estuary



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- | | |
|---------------------------------------------------------------------------------------|------------------------|
| | Mangrove swamp |
| | Mud |
| | Beach sand |
| ● | Subsurface Location |
| ■ | Live Material Location |
| ▲ | Surface Location |

II. MATERIALS AND METHODS

A. Sample collection

A total of 17 samples were gathered from various sites along the Savitri River (see Fig. 1), encompassing live, surface, and subsurface (shallow core) samples. To collect the subsurface samples, PVC pipes were utilized. Small cores were extracted using 4-inch diameter PVC pipes with a height of 10 cm, allowing for the retrieval of a significant amount of sediment. A cap was placed on the PVC pipe, with an arrow marked on it to indicate the upper section of the shallow core sample. In the laboratory, the subsurface cores were sectioned into three segments: top, middle, and bottom, each approximately 4 cm in length, and were processed and analyzed individually.

B. Foraminiferal analysis

For foraminiferal analysis, primarily entails the washing and screening of samples. The methodology applied is consistent for both surface and subsurface samples. The specific treatment employed is contingent upon the characteristics of the sediments from which microfossils are to be extracted. The samples predominantly comprised laterite and clay. The subsurface (shallow core) samples were uniformly divided into three segments of 4 cm each, categorized as top (0-4 cm), middle (4-8 cm), and bottom (8-12 cm). Each of these segments was treated independently.

Foraminiferal specimens were carefully selected and placed onto a microfossil slide with a 24-grid configuration for subsequent analysis. Their identification was conducted using a stereo microscope, aided by the methodologies outlined by Tappan and Loeblich Jr (1988).

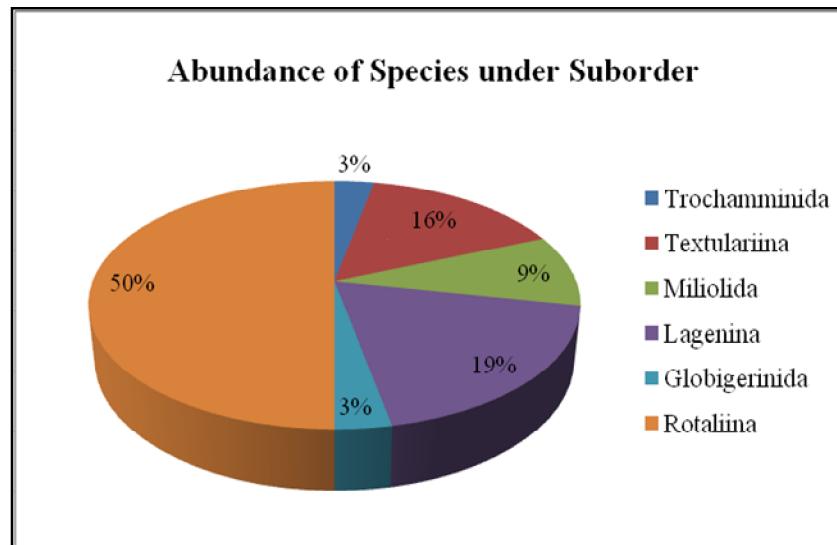


Fig. 2 - Abundance of Species under Suborder

C. Distribution

Table 1 - Distribution of foraminiferal species in different locations

A= Abundant (>20 specimens) M= Moderate (20-5 specimens) R= Rare (<5 specimens)

Sr. No.	Foraminifer Species	Velas	Bankot Jetty	Vesavi	Vesavi Jetty	Kante	Umroli
1	<i>Trochammina inflata</i>	M	-	M	R	M	R
2	<i>Ammodiscus</i> sp.	-	M	-	-	-	-
3	<i>Spiroplectammina</i> sp.	-	R	-	-	-	-
4	<i>Eggerelloides</i> sp.	-	M	-	-	-	-
5	<i>Ammobaculites</i> sp. A	-	R	-	-	-	-
6	<i>Ammobaculites</i> sp. B	-	R	-	-	-	-
7	<i>Milliammina fusca</i>	-	M	M	R	M	-
8	<i>Quinqueloculina poeyana</i>	R	A	-	M	-	R
9	<i>Quinqueloculina</i> sp. A	-	R	-	-	-	-
10	<i>Quinqueloculina</i> sp. B	-	R	-	-	-	-
11	<i>Hyalinonetrion gracillimum</i>	R	A	M	R	R	M
12	<i>Lagena semistriata</i>	-	A	A	-	-	A

13	<i>Lagena substriata</i>	R	A	M	-	R	-
14	<i>Lagena</i> sp.	-	R	-	-	-	-
15	<i>Favulina hexagona</i>	R	A	-	M	R	M
16	<i>Fissurina nudiformis</i>	M	M	R	-	M	-
17	<i>Bolivina striatula</i>	R	A	A	-	R	A
18	<i>Bolivina</i> sp.	-	M	-	-	-	-
19	<i>Bulimina marginata</i>	R	M	-	M	M	R
20	<i>Cassidelina</i> sp.	-	R	-	R	-	-
21	<i>Virgulinella pertusa</i>	-	A	R	-	M	M
22	<i>Nonionoides</i> sp.	-	R	-	-	-	-
23	<i>Nonionella</i> sp.	-	R	-	-	-	-
24	<i>Cancris auriculus</i>	-	A	R	-	-	M
25	<i>Rosalina globularis</i>	M	M	-	R	-	-
26	<i>Rosalina bradyi</i>	-	A	A	M	R	A
27	<i>Pararotalia</i> sp.	-	R	-	-	-	-
28	<i>Ammonia tepida</i>	-	A	A	R	-	A
29	<i>Elphidium advenum</i>	R	A	A	M	-	A
30	<i>Elphidium excavatum</i>	-	M	R	-	-	-
31	<i>Elphidium</i> sp.	-	R	-	-	-	-

III. SYSTEMATIC PALAEOONTOLOGY

Kingdom : PROTISTA Superclass : RHIZOPODA

Order : FORAMINIFERIDA

Suborder TROCHAMMINIDASaidova, 1981

Superfamily TROCHAMMINACEA Schwager, 1877

Family TROCHAMMINIDAE Schwager, 1877

Subfamily TROCHAMMININAE Schwager, 1877

Genus TROCHAMMINA Parker and Jones, 1859

Trochammina inflata Montagu, 1808

(Plate 1, Fig. 1)

Trochammina inflata (MONTAGU) Parker and Jones, 1859, p. 347.

Distinguishing Character: Test trochospiral; Whorls are inflated on the involute ventral side with a central depression.

Geological Range: Carboniferous to Holocene; cosmopolitan.

Ecology: epifaunal or infaunal, free; sediment; herbivore or detritivore; salinity 0-60; temperature 0-30°C; shelf 0-6000m; intertidal-abyssal (Murray, 2006).

Suborder TEXTULARIINA Delage and Herouard, 1896

Superfamily AMMODISACEA Reuss, 1862

Family AMMODISCIDAE Reuss, 1862

Subfamily AMMODISCINAE Reuss, 1862

Genus AMMODISCUS Reuss, 1862

Ammodiscus sp.

(Plate 1, Fig. 2)

Description: Test free, small, periphery rounded; proloculus not sharply defined, tubular second chamber planispirally coiled; spiral suture slightly overlapped, not depressed in appearance; wall agglutinated, smooth; aperture at open end of tubular second chamber.

Distinguishing Character: Sutures are overlapping and test is planispirally coiled.

Geological Range: Silurian to Holocene; cosmopolitan.

Distribution across Savitri Estuary: Very rare, only 2 specimens in Kante.

Superfamily SPIROPLECTAMMINACEA Cushman, 1927

Family SPIROPLECTAMMINIDAE Cushman, 1927

Subfamily SPIROPLECTAMMININAE Cushman, 1927

Genus SPIROPLECTAMMINIA Cushman, 1927

Spiroplectammina sp.

(Plate 1, Fig. 3)

Description: Test free, elongate, narrow, ovoid in section, margins broadly rounded, large early planispiral coil of few chambers followed by biserially arranged chambers, the coil commonly of greater breadth than the first few pairs of biserial chambers; wall agglutinated; aperture a low arch at the inner margin of the final chamber.

Distinguishing Character: Planispiral coiling followed by biserial arrangement of chambers.

Geological Range: Carboniferous to Holocene; cosmopolitan.

Superfamily TEXTULARIACEA Ehrenberg, 1838

Family EGGERELLIDAE Cushman, 1937

Subfamily EGGERELLINAE Cushman, 1937

Genus EGGERELLOIDES Haynes, 1973

Eggerelloides sp.

(Plate 1, Fig. 4)

Description: Test subfusiform, early stage trochospiral, later triserial; wall agglutinated, surface roughly finished; aperture in the centre of the slightly excavated apertural face.

Distinguishing Character: Early stage shows trochospiral coiling, later changes to triserial.

Geological Range: Holocene.

Ecology: infaunal, free; sand; detritivore; marine; temperate; shelf.

Superfamily HORMOSINACEA Haeckel, 1894

Family LITUOLIDAE de Blainville, 1827

Subfamily AMMOMARGINULININAE Podobina, 1978

Genus AMMOBACULITES Cushman, 1910

Ammobaculites sp. A

(Plate 1, Fig. 5)

Description: Test free, elongate portion close coiled, later uncoiling and rectilinear, rounded in section; wall agglutinated; aperture terminal, rounded.

Distinguishing Character: First portion close coiled then uncoiling and rectilinear.

Geological Range: L. Mississippian (Kinderhookian) to Holocene; cosmopolitan.

Ecology: infaunal, free; muddy sediment; detritivore; brackish-marine; temperate-tropical; brackish marshes and lagoons, inner shelf-upper bathyal (Murray, 2006).

Ammobaculites sp. B

(Plate 1, Fig. 6)



Description: Test free, elongate portion close coiled, later uncoiling; wall coarsely agglutinated; aperture terminal, elongated.

Distinguishing Character: First portion loosely coiled then uncoiling.

Geological Range: L. Mississippian to Holocene; cosmopolitan.

Superfamily RZEHAKINACEA Cushman, 1933

Family RZEHAKINIDAE Cushman, 1933

Genus MILIAMMINA Heron-Allen and Earland, 1930

Miliammina fusca

(Plate 1, Fig.7)

Distinguishing Character: Test elongate, ovate; aperture produced on a short neck.

Geological Range: L. Cretaceous to Holocene; cosmopolitan.

Ecology: infaunal – epifaunal, free; mud, silt, decaying vegetation; detritivore; brackish – hyper-saline; 0-30°C; marshes – upper bathyal (Murray, 2006).

Suborder MILIOLIDA Delage and Herouard, 1896

Superfamily MILIOLACEA Ehrenberg, 1838

Family MILIOLIDAE Ehrenberg, 1839

Subfamily QUINQUELOCUNINAE Cushman, 1917

Genus QUINQUELOCULINA d' Orbigny, 1826

Quinqueloculina poeyana d' Orbigny, 1826

(Plate 1, Fig. 8)

Quinqueloculina poeyana D' ORBIGNY, 1839, p. 191, pl. 11, figs. 25-27.

Distinguishing Character: Test quinqueloculine, ovate in outline and ornamented by closely spaced longitudinal striations.

Geological Range: Cretaceous to Holocene; cosmopolitan.

Ecology: epifaunal, free or clinging; plants or sediment; herbivore; marine – hypersaline; salinity 32-65; cold – warm; hypersaline lagoons, marine marsh and shelf, rarely bathyal (Murray, 2006).

Quinqueloculina sp. A

(Plate 1, Fig. 9)

Description: Test quinqueloculine, slightly elongated; three chambers are visible in side view, surface is ornamented by very light longitudinal ridges.

Distinguishing Character: Test quinqueloculine and Surface ornamentation.

Quinqueloculina sp. B

(Plate 1, Fig. 10)

Description: Test ovate in outline, early chambers quinqueloculine in both microspheric and megalospheric generations; three chambers are visible; wall calcareous, imperforate; aperture ovate.

Distinguishing Character: Ovate and imperforate test.

Suborder LAGENINA Delage and Herouard, 1896

Superfamily NUDOSARIACEA Ehrenberg, 1838

Family LAGENIDAE Reuss, 1862

Genus HYALINONETRION Patterson and Richardson, 1988

Hyalinonetria gracillimum Seguenza, 1862

(Plate 1, Fig. 11)

Amphorina gracillima SEGUENZA, 1862, p.51, pl. 1, figs. 37.

Distinguishing Character: Test unilocular; rhombic in outline, tapering equally on both ends of the chamber body; upper part with an elongated narrow neck; aboral end conical without any tubular extension of the test.

Geological Range: M. Oligocene to Holocene; cosmopolitan.

Subfamily ASIPHOLAGENINAE n. subf.

Genus LAGENA Walker and Jacob in Kanmocher, 1798

Lagena semistriata Williamson, 1848

(Plate 1, Fig.12)

Lagena striata var. *semistriata* WILLIAMSON, 1848, p. 14, pl. 1, figs. 9-10.

Description: Test unilocular, pyriform with an elongate neck that is ornamented by groove-like structure at the lowest part; surface smooth at the upper part of the chamber, but distinctly projecting downwards into crenulations.

Distinguishing Character: Test unilocular and groove-like structure at the base.

Geological Range: Jurassic to Holocene; cosmopolitan.

Lagena substriata Williamson, 1848

(Plate 1, Fig. 13)

Lagena substriata WILLIAMSON, 1848, p. 15, pl. 2, fig. 12.

Distinguishing Character: Longitudinal striae are present on whole test.

Geological Range: Jurassic to Holocene.

Lagena sp.

(Plate 1, Fig. 14)

Description: Test unilocular, globular to ovate, tapering at both ends but more sharp at lower end; wall calcareous, surface with longitudinal striae or costae; aperture terminal, rounded, produced on a short neck.

Distinguishing Character: Surface ornamented with longitudinal striations and aperture produced on a short neck.

Geological Range: Jurassic to Holocene.

Family ELLIPSOLAGENIDAE A. Silvestri, 1923

Subfamily OOLININAE Loeblich and Tappan, 1961

Genus FAVULINA Patterson and Richardson, 1988

Favulina hexagona Williamson, 1848

(Plate 1, Fig. 15)

Favulina hexagona (WILLIAMSON).

Distinguishing Character: Test Surface is ornamented with hexagonal pattern.

Geological Range: Miocene to Holocene; cosmopolitan.

Genus FISSURINA Reuss, 1850

Fissurina nudiformis McCulloch, 1977

(Plate 1, Fig. 16)

Fissurina nudiformis McCULLOCH, 1977, p. 118, pl. 59, figs. 1-5.

Distinguishing Character: Test unilocular, elongate-subglobular in outline with broadly rounded periphery; aperture terminal, tapering into flaps.

Geological Range: Cretaceous to Holocene; cosmopolitan.

Suborder ROTALIINADelage and Herouard, 1896

Superfamily BOLIVINACEA Glaessner, 1937

Family BOLIVINIDAE Glaessner, 1937

Genus BOLIVINAd' Orbigny, 1839

Bolivina striatulad' Orbigny, 1839

(Plate 1, Fig. 17)

Bolivina Striatula CUSHMAN, 1922, p.27, pl. 3, fig. 10.

Distinguishing Character: Test lamellar, elongate and lanceolate; chambers increasing slightly in size and biserial throughout; wall calcareous, hyaline and with very distinct pores; surface is distinctly ornamented by fine, longitudinal ribs over the initial 2/3 part of the test.

Geological Range: U. Cretaceous (Maastrichtian) to Holocene; cosmopolitan.

Bolivina sp.

(Plate 1, Fig. 18)

Description: Test rhomb shaped, tapering at both the ends gradually; wall calcareous; chambers are not clear due to sutural excavations (ridges), pronounced sutural excavations near the median line resulting in two thickened, elongated and slightly sinuate ridges; distinct pores are present throughout the test.

Distinguishing Character: All over the test ridges are present and its surface is ornamented by numerous pores.

Geological Range: U. Cretaceous to Holocene.

Superfamily BULIMINACEA Jones, 1875

Family BULIMINIDAE Jones, 1875

Genus BULIMINAd' Orbigny, 1826

Bulimina marginata' Orbigny, 1826

(Plate 1, Fig. 19)

Distinguishing Character: Test elongate-triangular, triserial throughout, chambers rapidly enlarging in height as added, strongly overlapping and slightly inflated; the lowermost acute margin of the chambers ornamented by short spines.

Geological Range: Paleocene to Holocene; cosmopolitan.

Ecology: infaunal, free; mud-fine sand; some species tolerate dysoxia; detritivore?; marine; cold-temperate; inner shelf-bathyal (Murray, 2006).

Superfamily TURRILINACEA Cushman, 1927

Family STAINFORTHIDAE Reiss, 1963

Genus CASSIDELINA Saidova, 1975

Cassidelina sp.

(Plate 1, Fig. 20)

Description: Test elongate, fusiform, biserial, periphery rounded; wall calcareous, perforated, smooth surface; aperture loop shaped; sutures slightly oblique, depressed inflated chambers in a twisted biserial arrangement, increasing rapidly in relative height as added, final pair occupying one third of the test length.

Distinguishing Character: Elongated test, fusiform and twisted biserial arrangement.

Geological Range: Pliocene to Holocene.

Superfamily FURSENKOINACEA Loeblich and Tappan, 1961

Family VIRGULINELLIDAE Loeblich and Tappan, 1984

Genus VIRGULINELLA Cushman, 1932

Vergulinella gunteri Cushman, 1932

(Plate 1, Fig. 21)

Virgulina floridana CUSHMAN, 1929, p. 54, pl. 9, figs. 7-10.

Distinguishing Character: Lower margin of every chamber shows various bridges or spine like projections and sutural openings between the sutured bridges.

Geological Range: Miocene to Holocene; cosmopolitan.

Superfamily NONIONACEA Schultze, 1854

Family NONIONIDAE Schultze, 1854

Subfamily NONIONINAE Schultze, 1854

Genus NONIONOIDES Saidova, 1975

Nonionoides sp.

(Plate 1, Fig.22)

Description: Test slightly asymmetrical and weakly trochospiral; chambers of nearly constant height but increasing rapidly in breadth to result in an auriculate test outline; sutures slightly depressed, gently curved, sides flattened, periphery rounded; wall calcareous, perforate with pustules or small spinules bordering the umbilical rim of the chambers in the vicinity of aperture.

Distinguishing Character: Wall perforated with small rounded pustules bordering the umbilical rim of the chambers in the vicinity of aperture.

Geological Range: Holocene.

Ecology: infaunal, free; muddy sediment; some species tolerate dysoxia; detritivore?; marine; temperate-warm (Murray, 2006).

Genus NONIONELLA Cushman, 1926

Nonionella sp.

(Plate 1, Fig. 23)

Description: Test free, calcareous, elongate, subovate, compressed, periphery rounded; chambers initially trochospirally organized, planispirally involute; chambers numerous, low, broad, increasing gradually in size; wall smooth; basal end broadly rounded; sutures generally radiate, curved centrally; last formed chamber with convex apertural face; aperture interiom marginal.

Distinguishing Character: Numerous chambers, radiating sutures and slightly asymmetrical in shape.

Geological Range: U. Cretaceous (Coniacian) to Holocene; cosmopolitan.

Ecology: infaunal, free; mud; some species tolerate dysoxia; detritivore?; marine; temperate-warm; shelf 10-1000m; shelf-upper bathyal (Murray, 2006).

Superfamily DISCORBACEA Ehrenberg, 1838

Family BAGGINIDAE Cushman, 1927

Genus CANCRIS de Montfort, 1808

Cancris auriculus Fichtel and Moll, 1798

(Plate 1, Fig. 24)

Cancris auriculus (Fichtel and Moll), 1798, p. 117, pl. 3, figs. 7-9, 16-18.

Distinguishing Character: Test in a flaring low trochospiral, auriculate in outline and lenticular in profile. Chambers are rapidly enlarging and periphery acute to carinate.

Geological Range: Eocene to Holocene; cosmopolitan.

Ecology: epifaunal, free; sediment; detritivore?; marine; temperate – subtropical; 50-150 m; shelf (Murray, 2006).

Family ROSALINIDAE Reiss, 1963

Genus ROSALINA d' Orbigny, 1826

Rosalina globularis d' Orbigny, 1826

(Plate 1, Fig. 25)

Rosalina globularis D' ORBIGNY, 1826, p. 271, pl. 13, figs. 1-4.

Distinguishing Character: Test lamellar, low trochospirally coiled, slightly planoconvex to concavoconvex; spiral sutures strongly oblique and slightly depressed, surface on this side densely and coarsely perforated. The final chamber is larger, encompassing nearly 1/3rd of circumference of the test.

Geological Range: Eocene to Holocene; cosmopolitan.

*Rosalina bradyi*Cushman, 1915

(Plate 1, Fig. 26)

Discorbis globularis var. *bradyi* CUSHMAN, 1915, p. 12, pl. 8, figs. 1a-c.

Distinguishing Character: Test lamellar, low trochospiral; spiral sutures are flush, thickened, imperforate and curved backwards at the periphery; surface densely and coarsely perforated on the side.

Geological Range: Eocene to Holocene; cosmopolitan.

Ecology: epifaunal, clinging or attached; hard substrates; herbivore?, omnivore; marine; temperate-warm; 0-100 m; lagoons; inner-mid shelf (Murray, 2006).

Superfamily ROTALIACEA Ehrenberg, 1839

Family ROTALIIDAE Ehrenberg, 1839

Subfamily PARAROTALIINAE Reiss, 1963

Genus PARAROTALIA Y. Le Calvez, 1949

Pararotalia sp.

(Plate 1, Fig. 27)

Distinguishing Character: Test in a low trochospiral coil, chambers flat to centrally elevated in the spiral side, commonly inflated and produced around the umbilicus; periphery carinate; sutures gently curved; wall calcareous.

Geological Range: U. Cretaceous (Coniacian) to Holocene; cosmopolitan.

Ecology: epifaunal, free; sand; herbivore; marine; warm; inner shelf (Murray, 2006).

Subfamily AMMONIINAE Saidova 1981

Genus AMMONIA Brünnich, 1772

*Ammonia tepida*Cushman, 1926

(Plate 1, Fig. 28)

Distinguishing Character: Test biconvex, low trochospiral coil of 2 to 3 evolutions, spiral side evolute, umbilical side involute; wall calcareous; 11 chambers are seen on dorsal side with few elongated and irregular ridges; sutures are straight or slightly curved and gently depressed.

Geological Range: L. Miocene to Holocene; cosmopolitan.

Ecology: infaunal, free; muddy sand; herbivore?; brackish, marine, hypersaline; warm temperature- tropical; shelf 0-50 m; brackish and hypersaline lagoons, inner shelf (Murray, 2006).

Family ELPHIDIIDAE Galloway, 1933

Subfamily ELPHIDIINAE Galloway, 1933

Genus ELPHIDIUM de Montfort, 1808

Elphidium advenum Cushman, 1922

(Plate 1, Fig. 29)

Elphidium adventum CUSHMAN, 1922, p. 25, pl. 10, figs. 1-2

Distinguishing Character: Test planispirally coiled, involute, slightly biconvex and symmetrical in profile view; peripheral margin angular to subacute, commonly keeled but occasionally non-keeled; 8-10 slightly inflated chambers; sutures curved backwards and crossed by 4-5 septal bridges.

Geological Range: L. Eocene to Holocene; cosmopolitan.



Ecology: (keeled species) epifaunal, free; sand, vegetation; herbivore; marine; temperate-warm; salinity 30-70; 0-50 m; inner shelf; (non-keeled species) infaunal, free; mud, sand; herbivore; salinity 0-70; brackish-hypersaline marshes and lagoons, inner shelf (upper bathyal, *Elphidium excavatum* only) (Murray, 2006).

Elphidium excavatum Terquem, 1875

(Plate 1, Fig. 30)

Distinguishing Character: Test planispirally coiled, peripheral margin subrounded, surface smooth with rounded periphery, pattern Geological Range: L. Eocene to Holocene; cosmopolitan.

Elphidium sp.

(Plate 1, Fig. 31)

Distinguishing Character: The ventral view of test is showing ridge-depression like arrangement around the centre.

Geological Range: L. Eocene to Holocene; cosmopolitan.

IV. CONCLUSION

The mangrove sediments of the Savitri Estuary have been found to contain dominantly calcareous foraminifera. The most prevalent native foraminiferal species within the mangrove ecosystem is *Trochammina inflata*. The foraminiferal community exhibits notably low diversity. Diversity peaks at the estuary's mouth and diminishes in the upstream areas. The documented collection suggests additional areas for investigation, particularly concerning changes in sea levels and the identification and classification of indigenous as Exotic fauna.

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REFERENCES

- [1] Abu-Zied, R.H., Basaham, A.S., El Sayed, M.A., 2013. Effect of municipal wastewater bottom sediment geochemistry and benthic foraminifera of two Red Seacoastal inlets, Jeddah, Saudi Arabia. Env. Ear. Sci. 68 (2), 451–469. doi: 10.1007/s12665-012-1751-7.
- [2] Badawi, A., El-Menhawy, W., 2016. Tolerance of benthic foraminifera to anthropogenic stressors from three sites of the Egyptian coasts. Egy. J. Aqu. Res. 42(1), 49–56. doi: 10.1016/j.ejar.2015.09.002.
- [3] Barbieri, R., Hohenegger, J., Pugliese, N., 2006. Foraminifera and environmental micropaleontology. Mar. Micropaleo. 61 (1–3), 1–3. doi: 10.1016/j.marmicro.2006.06.004, Issues.
- [4] Berkeley, A., Perry, C.T., Smithers, S.G., Horton, B.P., Taylor, K.G., 2007. A review of the ecological and taphonomic controls on foraminiferal assemblage development in intertidal environments. Ear. Sci. Rev. 83 (3–4), 205–230.
- [5] Bickert, T., & Wefer, G., 1999. South Atlantic and benthic foraminifer $\delta^{13}\text{C}$ deviations: implications for reconstructing the Late Quaternary deep-water circulation. Deep Sea Research Part II: Top. Stud. in Ocea., 46(1–2), 437–452.
- [6] Blainville, H. M. D. de., 1825–1827. Manuel de malacologie et de conchyliologie. Paris, Levrault 1-647 [1825], 649-664 + 109 pl. [1827].
- [7] Brünich, M.T., 1772. Zoologiae fundamenta paelectionibus academicis. Accomodata grunde I dyloeren. Transactions of the Linnean Society of London, 7:241.
- [8] Choi, J.U., An, S., 2012. High benthic foraminiferal diversity in polluted Busan NorthPort (Korea). J. Foram. Res. 42 (4), 327–339.
- [9] Cushman, J.A., 1910. A monograph of the foraminifera of the North Pacific Ocean. Pt. I - Astrorhizidae and Lituolidae. U.S. National Museum Bulletin, 71(1):1-134.
- [10] Cushman, J.A., 1915. A monograph of the foraminifera of the North Pacific Ocean, Part 5: Rotaliidae. Bulletin of the United States National Museum, 71(5):1-87.
- [11] Cushman, J.A., 1917. A monograph of the foraminifera of the North Pacific Ocean, Part 6: Miliolidae. Bulletin of the United States National Museum, 71(6):1-108.
- [12] Cushman, J. A., 1922. Shallow-water Foraminifera of the Tortugas region. Publications of the Carnegie Institution of Washington 311. Department of Marine Biology of the Carnegie Institution of Washington. Volume 17: 1-85.
- [13] Cushman, J.A., 1926. Foraminifera of the typical Monterey of California. Contributions from the Cushman Foundation for Foraminiferal Research, 2(3):53-69.
- [14] Cushman, J.A., 1927. An outline of a re-classification of the foraminifera. Contr. Contr. Lab. Foram. Res. 3 : 1-105.
- [15] Cushman, J. A., 1932. Notes on the genus Virgulina. Contributions from the Cushman laboratory for Foraminiferal Research. 8(1): 7-23.
- [16] Cushman, J.A., 1933. The foraminifera of the tropical Pacific collections of the "Albatross," 1899-1900, Part 2: Lagenida to Alveolinellidae. Bulletin of the United States National Museum, 161:1-79.
- [17] Cushman, J.A., 1937. A monograph of the foraminiferal family Valvulinidae. Special Publication Cushman Laboratory for Foraminiferal Research, 7:1-157.

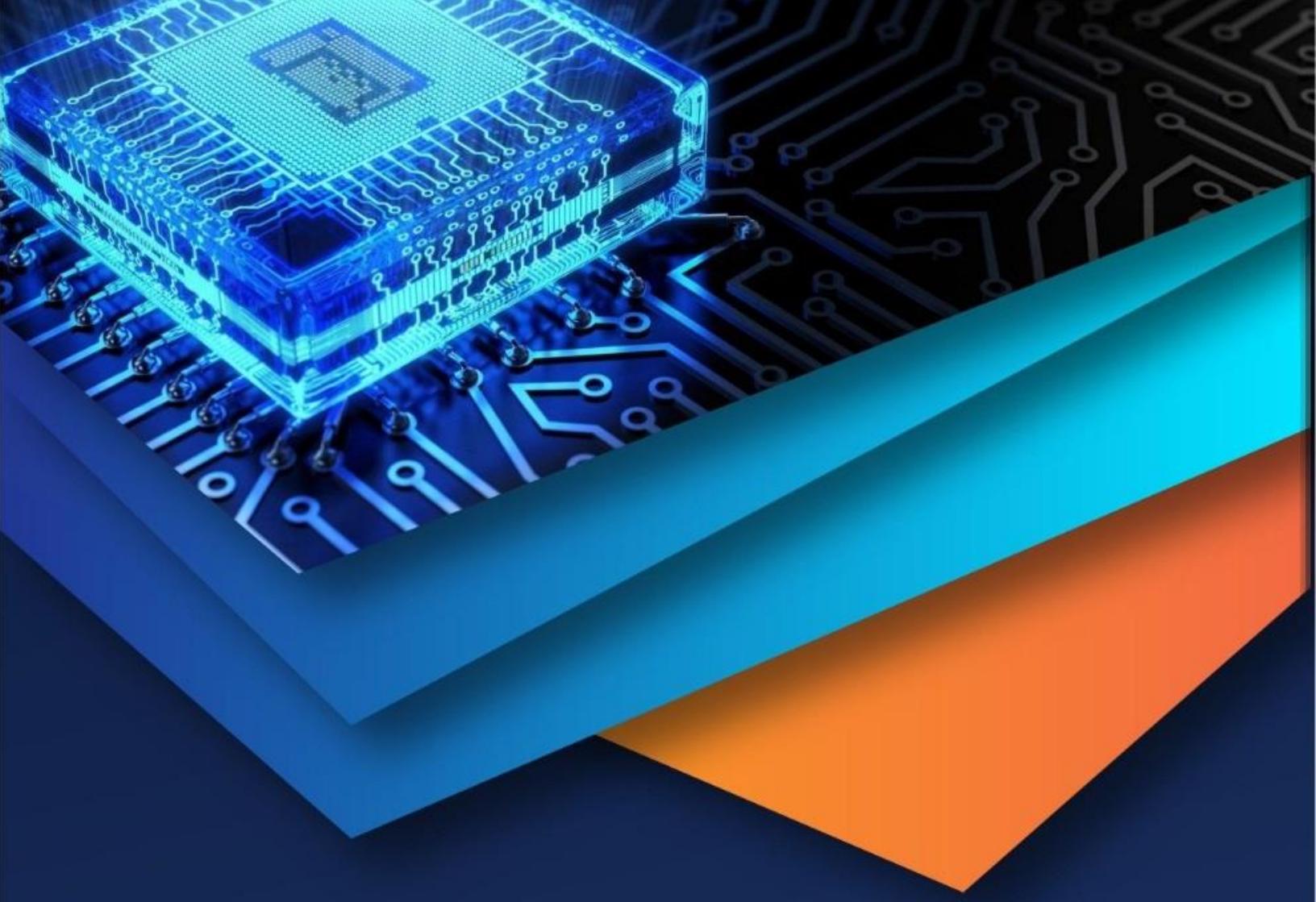
- [18] d'Orbigny, A., 1826. Tableau methodique de la classe des céphalopodes. *Annales des Sciences Naturelles*, 7:96-314.
- [19] de Montfort, D., 1808. *Conchyliologie Systematique et Classification Methodique des Coquilles*, 1. F. Schoell, Paris.
- [20] Debenay, J.P., Millet, B., Angelidis, M.O., 2005. Relationships between foraminiferal assemblages and hydrodynamics In The Gulf Of Kalloni, Greece. *J. Foram. Res.*35 (4).
- [21] Delage, Y. and Herouard, E., 1896. *Traité de Zoologie Concrete*. 1, La Cellule et les Protozoaires. Schleicher Frères, Paris.
- [22] Delage, Y.; Hérouard, E., 1896. *Traité de Zoologie Concrète. Tome I. La Cellule et les Protozoaires*. Schleicher Frères, Paris. 1-584.
- [23] Dimiza, M.D., Trianaphyllou, M.V., Portela, M., Koukousioura, O., Karageorgis, A.P., 2022. Response of living benthic foraminifera to anthropogenic pollution and metal concentrations in saronikos gulf (greece, eastern mediterranean). *Minerals*12 (5), 591.
- [24] Dong, S., Lei, Y., Li, T., Jian, Z., 2020. Response of benthic foraminifera to pH changes: community structure and morphological transformation studies from a microcosm experiment. *Mar. Micropaleo.* 156. doi: 10.1016/j.marmicro.2019.101819.
- [25] d'Orbigny, A., 1839. *Voyage dans l'Amérique Méridionale; Foraminifères*, 5. Levrault, Strasbourg, France.
- [26] Duplessy, J.C., Shackleton, N.J., Fairbanks, R.G., Labeyrie, L., Oppo, D., Kallel, N., 1988. Deepwater source variations during the last climatic cycle and their impact on the global deepwater circulation. *Paleoce 3* (3), 343-360.
- [27] Ehrenberg, C.G., 1839. Über die Bildung der Kreidefelsen und des Kreidemergels durch unsichtbare Organismen. *Abhandlungen der königlichen Akademie der Wissenschaften zu Berlin*, 1838:59-148.
- [28] Ehrenberg, G.C., 1838. Über dem blossen Auge unsichtbare Kalkthierchen und Kieselthierchen als Hauptbestandtheile der Kreidegebirge. *Bericht über die zur Bekanntmachung geeigneten Verhandlungen del Königlichen Preussischen Akademie der Wissenschaften zu Berlin*, 1838:192-200.
- [29] Fichtel, L.v. and Moll, J.P.C.v. 1798. *Testacea Microscopica, Aliaque Minuta ex Generibus Argonauta et Nautilus, ad Naturam picta et Descripta (Microscopische und andere kleine Schalthiere aus den Geschlechtern Argonauta und Schiffer)*. Camesina, Wien.
- [30] Galloway, J.J., 1933. *A Manual of Foraminifera*. Principia Press, Bloomington.
- [31] Glaessner, M.F., 1937. Die Entfaltung der Foraminiferenfamilie Buliminidae. *Problemy Paleontologii, Paleontologicheskaya Laboatoriya Moskovskogo Gosudarstvennogo Universiteta*, 2-3:411-422.
- [32] Haeckel, E., 1894. *Systematische Phylogenie. Entwurf eines Natürlichen Systems der Organismen auf Grund ihrer Stammesgeschichte. Systematische Phylogenie der Protisten und Pflanzen*, 1. Georg Reimer, Berlin.
- [33] Haynes, J. R., 1973. Cardigan Bay Recent Foraminifera (Cruises of the R. V. Antur, 1962-1964). *Bulletin of the British Museum (Natural History), Zoology. Supplement 4*, pp. 1-245.
- [34] Hemleben, C., Bijma, J., 1994. Foraminiferal population dynamics and stable carbon isotopes. In: *Carbon Cycling in the Glacial Ocean: Constraints on the Ocean's Role in Global Change: Quantitative Approaches in Paleoceanography*. Springer, Berlin Heidelberg, pp. 145-166.
- [35] Heron-Allen, E. and Earland, A., 1930. Some new foraminifera from the South Atlantic, Part 3. *Journal of Royal Microscopical Society*, 50:38-45.
- [36] Jones, T.R., 1875. In Griffith, J.W., and Henfrey, A. (eds.), *The Micrographic Dictionary*. van Voorst, London.
- [37] Kemp, A.C., Wright, A.J., & Cahill, N., 2020. Enough Is Enough, Or More IsMore? Testing The Influence of Foraminiferal Count Size on Reconstructions of Paleo-Marsh Elevation. In *Journal of Foram. Resea.* (Vol. 50, Issue3).
- [38] Le Calvez, Y., 1949. Révision des foraminifères Lutétiens du Bassin de Paris. Rotaliidae et familles affines. *Mémoires du Service de la Carte Géologique Détaillée de la France*, 2:1-54.
- [39] Li, T., Xiang, R., Li, T., 2014. Influence of trace metals in recent benthic foraminiferadistribution in the Pearl River Estuary. *Mar. Micropaleo.* 108, 13-27. doi: 10.1016/j.marmicro.2014.02.003.
- [40] Loeblich, A. R.; Tappan, H., 1984. Suprageneric Classification of the Foraminiferida (Protozoa). *Micropaleontology*. 30(1): 1-70. <https://doi.org/10.2307/1485456>.
- [41] Loeblich, A.R., Jr. and Tappan, H., 1961. Suprageneric classification of the Rhizopoda. *Journal of Paleontology*, 35(2):245-330.
- [42] Mackensen, A., Fu, D.K., Grobe, H., Schmiedl, G., 1993. Benthic foraminiferal assemblages from the eastern South Atlantic Polar Front region between 35 and 57 S: distribution, ecology and fossilisation potential. *Mar. Micropale.* 22 (1-2), 33-69.
- [43] McCulloch, I., 1977. Qualitative observations on Recent foraminiferal tests with emphasis on the eastern Pacific. University of Southern California, Los Angeles, California.
- [44] Minhat, F.I., Ghandhi, S.M., Ahzan, N.S.M., Haq, N.A., Manaf, O.A.R.A., Sabohi, S.M., Lee, L.H., Akhir, M.F., Abdullah, M.M., 2021. The occurrence and distribution of benthic foraminifera in tropical waters along the strait of malacca. *Front.Mar.Sci.* 8. doi: 10.3389/fmars.2021.647531.
- [45] Montagu, G., 1808. Supplement to *Testacea Britannica* with Additional Plates. Woolmer, Exeter. v + 183 pp., pl. 17-30.
- [46] Murray, J.W. 1991. *Ecology and palaeoecology of benthic foraminifera*: longman Scientific and Technical. Harlow, England, 2000, 244–245.
- [47] Murray, J.W., 2006. *Ecology and Applications of Benthic Foraminifera*. Cambridge University Press, New York, 1-426. <https://doi.org/10.1017/S0016756808004676>.
- [48] Nomaki, H., Ogawa, N.O., Ohkouchi, N., Suga, H., Toyofuku, T., Shimamura, M., Nakatsuka, T., Kitazato, H., 2008. Benthic foraminifera as trophic links between phytopdetritus and benthic metazoans: carbon and nitrogen isotopic evidence. *Mar.Eco.Progress Series* 357, 153–164. doi: 10.3354/meps07309.
- [49] Parker, W. K.; Jones, T. R. 1859. On the nomenclature of the Foraminifera. II. On the species enumerated by Walker and Montagu. *Annals and Magazine of Natural History*. (3) 4 (23): 333-351.
- [50] Patterson, R.T. and Richardson, R.H., 1988. Eight new genera of unilocular foraminifera (Lagenidae). *Transactions of the American Microscopical Society*, 107:240-258. <https://doi.org/10.2307/3226501>.
- [51] Podobina, V.M., 1978. *Sistematička i filogenija Gaplofragmiidej*. Tomsk Universitet, Tomsk. (In Russian).
- [52] Reiss, Z., 1963. Reclassification of perforate foraminifera. *Bulletin of the Geological Survey of Israel*, 35:1-111.
- [53] Reuss, A.E., 1850. Neues Foraminiferen aus den Schichten des österreichischen Tertiärbeckens. *Denkschriften der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Classe*, 1:365-390.
- [54] Reuss, A. E., 1862. Entwurf einer systematischen Zusammenstellung der Foraminiferen. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe. Abt. 1, Mineralogie, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*. (1861) 44(1).355-396.

- [55] Saidova, K.M., 1975. Bentosnye Foraminifery Tikhogo Okeana (Benthonic foraminifera of the Pacific Ocean). Institut Okeanologii P.P. Shirshova, Akademiya Nauk SSSR, Moscow.
- [56] Saidova, K. M., 1981. On the current state of the system of supraspecific taxa of Cenozoic benthic foraminifera. Akademiya Nauk SSSR. 1-73.
- [57] Schafer, C.T., Collins, E.S., Smith, J.N., 1991. Relationship of Foraminifera and thecamoebi distributions to sediments contaminated by pulp mill effluent:saguenay Fiord, Quebec, Canada. Mar. Micropale. 17 (3-4), 255-283. doi: 10.1016/0377-8398(91)90016-Y.
- [58] Schmiedl, G., Leuschner, D.C., 2005. Oxygenation changes in the deep western Arabian Sea during the last 190,000 years: productivity versus deepwater circulation. Paleocean 20 (2).
- [59] Schultze, M.S., 1854. Über den Organismus der Polythalamien (Foraminiferen), nebst Bemerkungen über die Rhizopoden im Allgemeinen. Wilhelm Engelmann, Leipzig.
- [60] Schwager, C., 1877. Quadro del proposto sistema di classificazione dei foraminiferi con guscio. Bollettino R. Comitato Geologico d'Italia. 8: 18-27.
- [61] Seguenza, G., 1862. Dei terreni Terziarii del Distretto di Messina, Parte II, Descrizione dei Foraminiferi Monothalamici Delle Marne Mioceniche del Distretto di Messina. Capra, T., Messina.
- [62] Silvestri, A., 1923. Lo stipe della Elisoforme e le sue affinità. Memorie della Pontificia Accademia della Scienze, Nuovi Lincei, 2(6):231-270.
- [63] Tappan, H., Loeblich, A.R., 1988. Foraminiferal Genera and Their Classification. VanNostrand Reinhold.
- [64] Terquem, O., 1875. Essai sur le classement des animaux qui vivent sur la plage et dans les environs de Dunquerque. Memoirs de la Society Dunkerquoise, 19:405-447.
- [65] Walker, G. and Jacob, E., 1798. An arrangement and description of minute and rare shells, p. 629-645. In Adams, G. and Kanmacher, F. (eds.), Essays on the Microscope. Dillon and Keating, London.
- [66] Williamson, W.C., 1848. On the Recent British species of the genus *Lagena*. The Annals and Magazine of Natural History, 2, 1:1-20. <https://doi.org/10.1080/03745485809494465>.
- [67] Wukovits, J., Oberrauch, M., Enge, A.J., Heinz, P., 2018. The distinct roles of two intertidal foraminiferal species in phytodetrital carbon and nitrogen fluxes - results from laboratory feeding experiments. Biogeoscience 15 (20), 6185-6198. doi: 10.5194/bg-15-6185-2018.





Plate 1-(1) *Trochammina inflata* (Montagu, 1808) (Scale bar = 100 µm). (2) *Ammodiscus* sp. (Scale bar = 50 µm). (3) *Spiroplectammina* sp. (Scale bar = 100 µm). (4) *Eggerelloides* sp. (Scale bar = 100 µm). (5) *Ammobaculites* sp. A (Scale bar = 200 µm). (6) *Ammobaculites* sp. B (Scale bar = 100 µm). (7) *Milliammina fusca* (Scale bar = 50 µm). (8) *Quinqueloculina poeyana* (d' Orbigny, 1826) (Scale bar = 50 µm). (9) *Quinqueloculina* sp. A (Scale bar = 50 µm). (10) *Quinqueloculina* sp. B (Scale bar = 50 µm). (11) *Hyalinonetrion gracillimum* (Seguenza, 1862) (Scale bar = 100 µm). (12) *Lagena semistriata* (Williamson, 1848) (Scale bar = 100 µm). (13) *Lagena substriata* (Williamson, 1848) (Scale bar = 20 µm). (14) *Lagena* sp. (Scale bar = 50 µm). (15) *Favulina hexagona* (Williamson, 1848) (Scale bar = 50 µm). (16) *Fissurina nudiformis* (McCulloch, 1977) (Scale bar = 50 µm). (17) *Bolivina striatula* (d' Orbigny, 1839) (Scale bar = 100 µm). (18) *Bolivina* sp. (Scale bar = 20 µm). (19) *Bulimina marginata* (d' Orbigny, 1826) (Scale bar = 50 µm). (20) *Cassidelina* sp. (Scale bar = 100 µm). (21) *Vergulinella gunteri* (Cushman, 1932) (Scale bar = 50 µm). (22) *Nonionoides* sp. (Scale bar = 100 µm). (23) *Nonionella* sp. (Scale bar = 50 µm). (24) *Cancris auriculus* (Fichtel and Moll, 1941) (Scale bar = 50 µm). (25) *Rosalina globularis* (d' Orbigny, 1826) (Scale bar = 100 µm). (26) *Rosalina bradyi* (Cushman, 1915) (Scale bar = 50 µm). (27) *Pararotalia* sp. (Scale bar = 100 µm). (28) *Ammonia tepida* (Brünnich, 1771) (Scale bar = 50 µm). (29) *Elphidium advenum* (Cushman, 1930) (Scale bar = 50 µm). (30) *Elphidium excavatum* (Terquem, 1875) (Scale bar = 50 µm). (31) *Elphidium* sp. (Scale bar = 50 µm).



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