



iJRASET

International Journal For Research in
Applied Science and Engineering Technology



INTERNATIONAL JOURNAL FOR RESEARCH

IN APPLIED SCIENCE & ENGINEERING TECHNOLOGY

Volume: 7 Issue: I Month of publication: January 2019

DOI: <http://doi.org/10.22214/ijraset.2019.1071>

www.ijraset.com

Call: ☎ 08813907089

E-mail ID: ijraset@gmail.com

The Effect of Exogenous Application of GA₃ to Mitigate the Salt Induced Damages in Rice Cultivars during Germination

Dr. K. Krishna

Assistant Professor, P.G. Department of Botany, Yuvaraja's College, (Mysore University constituent autonomous college)
University of Mysore. Mysore. Karnataka.

Abstract: *In the present study the two paddy cultivars were selected to examine the effect of salinity, GA₃ and the effect of GA₃ on the salinity stressed seeds, the tests were conducted at different concentrations. The salinity decreased the germination percentage, vigour index and seedling length and chlorophyll content. GA₃, combination of GA₃ and NaCl treated seeds showed increased germination percentage in both the cultivars, but the decrease in germination percentage due to salinity was more in Jyothi cultivar compared to the BPT Super cultivar. GA₃ played the salinity attenuating role, Jyothi cultivars seems to be more salt sensitive when compared to the BPT Super cultivar. The Chlorophyll a, Chlorophyll b and total chlorophyll content was decreased in salinity in both the varieties, but a gradual increase was found in GA₃ treatment and also combination of NaCl and GA₃ treatments. The decrease in chlorophyll content under salinity was more in Jyothi cultivar when compared to the BPT Super cultivar.*

Keywords: *Rice, NaCl, GA₃, seed germination, seedling length, chlorophyll.*

I. INTRODUCTION

Rice (*Oryza sativa* L.) is a salt sensitive monocot and widely grown crop in tropical and subtropical regions. It is one of the main staple foods for nearly two thirds of the population of the world (Roy *et al.*, 2012). High rice consumption, degradation of soil and water quality around the globe has focussed urgent attention to understand the response of this important crop towards abiotic stresses. Salinity is estimated that over 800 million hectares of land in the world are affected (Munns, 2005; Kumar *et al.*, 2010). Salt stress in soil is one of the major stresses especially in arid and semi arid regions and can severely limit plant growth and productivity by reducing osmotic potential, ion toxicity creation, uptake disarrangement and ion imbalance and can cause disorders in enzyme activities of membrane and metabolic activities in plants (Gorham, 1993; Hasegawa *et al.*, 2000; Basu *et al.*, 2002; Murphy *et al.*, 2003; Islam *et al.*, 2008). These activities could affect morphological parameters and plant growth and will reduce vegetative growth (Linghe and Shannon, 2000; Sairam and Tyagi, 2004). Chlorosis is common morphological and physiological characteristic of mulberry in response to salt stress (Harinasut *et al.*, 2000). Chlorophyll is a good indicator of plant nutrient stress during growing period and content of chlorophyll in the leaves indicates the growth of crops. Synthesis and integrity of chlorophyll level may vary due to salt stress (Santo, 2004; Rout *et al.*, 1997). Chlorophyll content of salt stressed rice can be described as functions of the sodium content (Yeo and Flowers, 1983). Excess of NaCl to plants involves changes in their morphology, physiology and metabolism (Hilal *et al.*, 1998; Rahman *et al.*, 2008) and consequently reducing plant dry weight (Zeng and Shannon, 2000; Razzaque *et al.*, 2009) and dry matter production (Mansour and Salam, 2007) and ultimately its effect on crop yield (Shannon *et al.*, 1998; Jamil *et al.*, 2010; Osakabe *et al.*, 2011). Several strategies have been proposed to alleviate the degree of cellular damage caused by abiotic stress and to improve crop salt tolerance. Among them exogenous application of compatible osmolytes such as proline, glycinebetaine, trehalose, auxin, gibberellins, (Kim *et al.*, 2006; Rahdari *et al.*, 2012), Methyl Jasmonate, Sucrose (Siringam *et al.*, 2012), Spermidine (Saleethong *et al.*, 2016) had gained considerable attention in mitigating the effect of salt stress. Plant growth regulators and other groups of chemicals have been used to treat rice plants exogenously at various growth stages to increase salt tolerance by alleviating salt induced damages and led to improved growth and productivity (Roychoudhury *et al.*, 2011; Plaut *et al.*, 2013). A few studies have however, demonstrated the ability of GA₃ to overcome adverse effects of NaCl stress (Chakraborti and Mukherji, 2003). GA₃ basically stimulated the inter node elongation, and also controls the various aspects of seed germination, mobilization of reserves, floral initiation, sex determination and fruit set. The present work was conducted to examine whether NaCl inhibited seed germination, growth and synthesis of chlorophyll content and to determine the influence of GA₃ when exposed to NaCl stress.

II. MATERIALS AND METHODS

Paddy seed sample of cultivars Jyothi and BPT Super were procured from VC Farm, Regional Agricultural Research Station, University of Agriculture Science, Mandya, Karnataka. Seeds of uniform size were selected and surface sterilized using 0.01% of mercuric chloride for two minutes. The seeds were washed thoroughly with distilled water for 4-5 times and soaked for 24 hours in distilled water (control) and different concentrations of GA₃ i.e 100ppm and 200ppm and in NaCl i.e. 200mM and 300mM and in combination of NaCl and GA₃. The germination studies were carried as per International Seed Testing Association (2009). Five sets of each concentration of GA₃, NaCl and combination of both were maintained along with control. The seeds were allowed to germinate for 14days as per ISTA, 2009 and then analysed for morphological and biochemical parameters.

The test was conducted by the between paper towel method recommended (ISTA, 2009). Hundred seeds of each cultivar were placed on craft paper saturated with known concentration of sodium chloride and GA₃. The number of seeds germinated in each treatment was counted on 5th day of germination and total germination percentage was calculated. On the 14th day root length, shoot length and seedling length was measured and The seedling vigour index was calculated by using the formula proposed by Abdul Baki and Anderson (1973) and expressed as seedling vigour index (SVI) = % of Germination X Mean seedling length. The chlorophyll content viz., Chlorophyll-a and Chlorophyll-b and the total chlorophyll were estimated as per the method of Arnon (1949).

III. RESULTS AND DISCUSSION

A. Seed Germination

Rice is one of the most important cereal crops all over the world with exceptional agricultural and economic importance as a staple food for more than 50% population worldwide. Asian farmers produce more than 90% of the total rice. The test was conducted to study the germination percentage of two rice cultivars subjected to salinity stress. As per investigation there was decrease in germination percentage under saline treatment and there was increase in the germination percentage under GA₃ treatment GA₃ alleviated the negative effect of NaCl.

B. Effect of Salinity on Germination

Salinity is a common environmental stress seriously affecting on crop growth and crop yield in many regions, particularly crops in arid and semi-arid regions (Jamil *et al.*, 2010; Osakabe *et al.*, 2011). It is estimated that over 800 million hectares of land in the world are affected by salinity (Munns, 2005; Kumar *et al.*, 2010).

The salinity decreased germination percentage, vigour index and seedling length (table 1). Salinity induces numerous disorders in seeds during germination, it reduces the imbibitions of water because of lower osmotic potential of the medium, (Akbar and Ponnampurna, 1982, Almansouri, *et al.*, 2001). Salinity causes mineral imbalance and toxicity in the environment, so that the mineral imbalance and toxicity of the saline environment often affects the structure and chemical composition of bilayer lipid membrane of the seed. It also affects the membrane selective ability to transport of the solutes and ions inward and also becomes leaky to solutes they contain and hence the germination percentage is affected (Cushman, *et al.*, 2001).

Salinity decreased the germination percentage, magnitude of reduction increased with increasing salinity stress (Hakim, *et al.*, 2009 and Mirza Hasamuzzaman *et al.*, 2009 and Anbumalarnathi, *et al.*, 2013). Salinity of soil and water is a major environmental stress that drastically affects on crop productivity in term of growth and yield, all over the world (Zhu, 2001).

C. Effect of GA₃ on Germination

Gibberellin increased the cell division and elongated the cells and it effected positively on the germination (Parvanehrahdari *et al.*, 2015). The interaction of GA₃ and NaCl concentrations was no significant, but optimal concentration of GA₃ treatment was slightly increased germination percentage. The exogenous application of plant growth regulator like gibberellins (Afzal *et al.*, 2006), reduce the adverse effects of salt stress and also improves germination, growth, development and seed yields and yield quality (table 1).

D. Effect of salinity and GA₃ on germination

The test was conducted to study the germination percentage of two rice cultivars subjected to salinity stress (table 1). As per our investigation there was decrease in germination percentage under saline treatment due to osmotic potential of salinity on imbibitions of seeds was the main cause for reduced germination (Akbar and Ponnampurna 1982; Almansouri, *et al.*, 2001) and increase in the germination percentage under GA₃ treatment. The NaCl treated seeds were treated with GA₃ the germination percentage was increased, hence GA₃ alleviated the negative effect of NaCl.

E. Effect of Salinity and GA₃ on Vigour Index

The vigour index was decreased under saline treatment in both the varieties. There was increase in vigour index under GA₃ treatment. The combination of GA₃ and NaCl also showed increase in the vigour index. Vigour index of Jyothi cultivar was most affected under salinity when compared to the BPT Super cultivar. Vigour index is directly dependent on germination percentage and seedling length. Germination percentage and the seedling length of saline treated seeds were decreased. Therefore the vigour index was also decreased. Under GA₃ treatment and combination of GA₃ and NaCl seeds showed increase in germination percentage and seedling length and hence there was increase in the vigour index too.

Table 1: Effect of different concentration of NaCl, GA₃ and combination of NaCl and GA₃ on germination percentage (%) and vigour index of Jyothi and BPT Super paddy cultivars on 5th day.

Cultivar	Parameters	Control	NaCl 200mM	NaCl 300mM	GA ₃ 100ppm	GA ₃ 200ppm	GA ₃ 100ppm + NaCl 200mM	GA ₃ 200ppm + NaCl 300mM
Jyothi	Germination %	96.66 ^{ab}	92.00 ^c	87.33 ^d	96.00 ^{abc}	98.66 ^a	94.00 ^{bc}	96.66 ^{ab}
	Vigour index	1985.2 ^{bcd}	1545.4 ^{de}	1313.1 ^e	2218.5 ^{abc}	2669.2 ^a	1808.6 ^{cde}	2457.5 ^{ab}
BPT Super	Germination %	92.66 ^{bc}	90.00 ^{cd}	86.66 ^d	96.00 ^{ab}	98.66 ^a	92.00 ^{bc}	95.33 ^{ab}
	Vigour index	1395.9 ^{bc}	1162.4 ^{cd}	1040.3 ^d	1650.5 ^{ab}	1941.7 ^a	1440.6 ^{bc}	1715.4 ^{ab}

Means followed by the same letter within a row are not significantly different as indicated by

Scheffe ($P \leq 0.05$) significant at $P \leq 0.001$.

F. Effect Of Salinity On Seedling Length

The seedling length was decreased under salinity and increase in the seedling length was found in GA₃ treatment. Combination of NaCl and GA₃ also showed increase in seedling length in both Jyothi and BPT Super cultivars. Seedling length was less decreased in BPT Super cultivar under saline treatment compared to the Jyothi cultivar (table 2). The gradual decrease in root length with the increase in salinity was observed might be due to more inhibitory effect of NaCl salt on the root growth (Rahman, *et al.*, 2001).

The concentration of NaCl modifies the metabolic activities of the cell wall causing the deposition of various materials which limit the cell wall elasticity, secondary cell walls become rigid and consequently the turgor pressure efficiency in cell enlargement is decreased, it affects the growth of seedling (Ali *et al.*, 2004). Due to salinity seed fertility was damaged hence the seedling length was decreased (Thanaphol *et al.*, 2012). Salinity led to the reduction in shoot and root length of rice varieties and the magnitude of reduction increased with increasing salinity stress (Hakim, *et al.*, 2009; Anbumalaramathi, *et al.*, 2013 and Mirza Hasamuzzaman *et al.*, 2009).

G. Effect of GA₃ on Seedling Length

GA₃ treatment and combination of GA₃ and NaCl increased the germination percentage of both the cultivars (table 2) due to increases the cell division, elongation of cells and helps in the increase of seedling length (Parvanehrahdari *et al.*, 2015). GA₃ enhanced the growth by forming new cells in the intercalary meristem (Khadija *et al.*, 2013).

Table 2: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on root, shoot and seedlings length (cm) of Jyothi and BPT Super Paddy cultivars on 14th day.

Cultivar	Parameters	Control	NaCl 200mM	NaCl 300mM	GA ₃ 100ppm	GA ₃ 200ppm	GA ₃ 100ppm + NaCl 200mM	GA ₃ 200ppm + NaCl 300mM
Jyothi	Root length	8.87 ^b	7.59 ^{bc}	6.92 ^c	11.32 ^a	12.36 ^a	8.39 ^{bc}	11.06 ^a
	Shoot length	11.65 ^{abc}	9.21 ^c	8.13 ^c	11.80 ^{abc}	14.69 ^a	10.84 ^{bc}	14.36 ^{ab}
	Seedling length	20.62 ^{bcd}	16.94 ^{de}	15.12 ^e	23.16 ^{abc}	27.14 ^a	19.30 ^{cde}	25.46 ^{ab}
BPT Super	Root length	6.71 ^{cd}	5.77 ^{de}	5.01 ^e	7.66 ^{bc}	8.92 ^a	6.88 ^{bcd}	8.38 ^{ab}
	Shoot length	8.33 ^a	7.14 ^a	7.00 ^a	12.06 ^a	10.75 ^a	8.76 ^a	9.61 ^a
	Seedling length	15.15 ^{bcd}	13.01 ^{cd}	12.07 ^d	17.29 ^{ab}	19.74 ^a	15.75 ^{bc}	18.08 ^{ab}

Means followed by the same letter within a row are not significantly different as indicated by Scheffe

($P \leq 0.05$) significant at $P \leq 0.001$.

H. Chlorophyll

- 1) *Effect of salinity on chlorophyll*: The chlorophyll content was decreased under saline condition as the salinity increased (table 3) due to increased activity of the chlorophyll degrading enzyme chlorophyllase (Reddy *et al.*, 1986), ion accumulation and functional disorders observed during opening and closing of stoma under salinity stress (Seemann and Critchley, 1985; Khalehi *et al.*, 2012; Nawaz *et al.*, 2010). The reason for the decrease of chlorophyll content under salt conditions is the rapid maturity of leaves (Yeo, *et al.*, 1991). The depletion of chlorophyll under salinity may be the result of the hang-up of chlorophyll biosynthesis following an increase in ethylene production brought by the elevated NaCl content (Khan, 2003). Chlorophyll a, chlorophyll b, total chlorophyll content was decreased under salinity (Mohammad Reza Amirjani, *et al.*, 2011, Khadija, *et al.*, 2013, and Abhilash Joseph, *et al.*, 2014).
- 2) *Effect Of Ga₃ On Chlorophyll*: GA₃ played significant role in salinity alleviating and increased the chlorophyll content in the saline treated seeds (table 3). The positive effect of GA₃ was constant by having more chlorophyll than stressed conditions (Khadija, *et al.*, 2013). The GA₃ generated sweetening of ultra-structural morphogenesis of plastids coupled with the retention of chlorophyll and delay of senescence caused by GA₃ (Arteca, 1997). The salinity reduces in chlorophyll but GA₃ raised significantly (table 3).

Table 3: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on chlorophyll a, chlorophyll b and total chlorophyll (mg/g) of Jyothi and BPT Super paddy cultivars on 14th day.

Cultivar	Parameters	Control	NaCl 200mM	NaCl 300mM	GA ₃ 100ppm	GA ₃ 200ppm	GA ₃ 100ppm + NaCl 200mM	GA ₃ 200ppm + NaCl 300mM
Jyothi	Chl-a	0.028 ^d	0.010 ^e	0.008 ^e	0.032 ^{cd}	0.075 ^b	0.036 ^c	0.089 ^a
	Chl-b	0.009 ^d	0.007 ^d	0.006 ^d	0.027 ^b	0.044 ^a	0.008 ^d	0.019 ^c
	Total chl	0.070 ^d	0.026 ^e	0.023 ^e	0.084 ^c	0.192 ^b	0.089 ^c	0.219 ^a
BPT Super	Chl-a	0.030 ^c	0.027 ^c	0.027 ^c	0.036 ^{bc}	0.041 ^{bc}	0.038 ^{bc}	0.060 ^b
	Chl-b	0.007 ^{bc}	0.005 ^{cd}	0.003 ^d	0.01 ^{ab}	0.013 ^a	0.008 ^{bc}	0.009 ^b
	Total chl	0.073 ^{de}	0.060 ^e	0.060 ^e	0.086 ^{cd}	0.103 ^b	0.093 ^{bc}	0.151 ^a

Means followed by the same letter within a row are not significantly different as indicated by Scheffe ($P \leq 0.05$) significant at $P \leq 0.001$.

IV. CONCLUSION

In our present study the two paddy cultivar were selected to examine the effect of salinity, GA₃ and the effect of GA₃ on the salinity stressed seeds and the tests were conducted at different conditions with different concentrations. The various aspects were investigated. The salinity decreased germination percentage, vigour index and seedling length, GA₃ treatment and combination of GA₃ and NaCl increased the germination percentage of both the cultivars, but the decrease in germination percentage due to salinity was more in Jyothi cultivar compared to the BPT Super cultivar. GA₃ played the salinity attenuating role and Jyothi cultivars seems to be more salt sensitive when compared to the BPT Super cultivar. The chlorophyll content was decreased under saline condition as the salinity increased. GA₃ played significant role in salinity alleviating and increased the chlorophyll content in the saline treated seeds. The salinity alleviating role of GA₃ was observed and BPT Super cultivars chlorophyll content was less affected when compared to the Jyothi cultivar hence the BPT super cultivar is more tolerant compared to the Jyothi cultivar.

REFERENCE

- [1] Abdul-baki, B.A.A., and J.D. Anderson. 1973. Relationship between decarboxylation of glutamic acid and vigor in soybean seed. Crop Sci. 13:222-226.
- [2] Abhilash Joseph E, V.V. Radhakrishnan, K.T. Chandramohan and K.V. Mohanan.2014. Reduction of major photosynthetic pigments under salinity stress in some native rice cultivars of North Kerala, India. International Journal of Recent Scientific Research. 5: 1602- 1611.
- [3] Afzal I, S.M.A. Faoq, M. Nawaz 2006. Alleviation of salinity stress in spring wheat by hormonal priming with ABA, Salicylic acid and ascorbic acid. Int J. Agric. Biol. 8: 23-38.
- [4] Akbar, M. And F.N. Ponnampuruma, 1982. Saline soils of South and Southeast Asia as a potential rice lands. Rice Research Strategies for the Future. IRRI Los Banos, Laguna, Philippines.
- [5] Ali Y, Z. Aslam, M.Y. Ashraf and G.R. Tahir. 2004. Effect of salinity on chlorophyll concentration, leaf area, yield and yield components of rice genotypes grown under saline environment. International Journal of Environmental Science & Technology. 3: 221-225.

- [6] Almansouri, M., Kinet, M., Lutts, S. (2001), Effect of salt and osmotic stresses on germination in durum wheat (*Triticum durum* Desf.). *Plant and Soil*, 231, 243-254.
- [7] Anbumalaramathi J & Preeti Mehta. 2013. Effect of Salt Stress on Germination of indica Rice Varieties. *EJBS* 6 (1): 1-6.
- [8] Arnor .D.I. 1949. Copper enzymes in isolated chloroplasts. Poly phenoloxidase in *Beta vulgaris*. *Plant Physiol.* 24: 1-15.
- [9] Arteca, R.N. 1997. *Plant Growth Substances. Principles and Applications*. CBS Publishers, New Delhi.
- [10] Basu S., gangopadhyay G. and Mukherjee B. 2002. Salt tolerance in rice in vitro: implication of accumulation of Na^+ , K^+ and proline. *Plant Cell Tiss. Org. Cult.* 69:55-64.
- [11] Chakrabarti N, Mukherji S 2003. Alleviation of NaCl stress by pretreatment with phytohormones in *Vigna radiata*. *Biol. Plantarum* 46: 589-594.
- [12] Cushman, T.C. 2001. Osmoregulation in plants: Implications for agriculture. *American Zoologist*, 41(4): 758-769.
- [13] Gorham, J. 1993. *Genetics and physiology of enhanced of plant mineral nutrition*. Powal and Allen (ed), Kluwer Academic Publisher Dodrecht,. The Netherlands.
- [14] Hakim M. A., A. S. Juraimi, M. Begum, M. M. Hanafi, Mohd R Ismail and A. Selamat. 2009. Effect of salt stress on germination and early seedling growth of rice (*Oryza sativa* L.). *African Journal of Biotechnology*. 9(13): 1911-1918.
- [15] Harinasut P., S. Srinunaka. S. Pitukchaisopola and R. Charoensatapom. 2000. Mechanisms of adaptation to increasing salinity of mulberry: Proline content and ascorbate peroxidase activity in leaves of multiple shoots. *Sci. Asia*, 2006.
- [16] Hasegawa, P.M., R.A. Bressnan, J.K. Zhu and H.J Bohnert. 2000. Plant cellular and molecular responses to high salinity. *Annual Review of Plant Physiology and Plant Molecular Biology* 51: 463-499.
- [17] Hilal M., Zenoff A.M., Ponessa G., Moreno H. And Massa E.M. 1998: Saline stress alters the temporal patterns of xylem differentiation and alternative oxidative expression in developing soybean roots. *Plant Physiol.* 11: 695-701.
- [18] Islam M.S., Hur J.H. and Wang M.H. 2008. The influence of abiotic stresses on expression of zinc finger gene in rice. *Russ J. Pl. P.* 56:695-701.
- [19] ISTA.2009. International seed testing association, News Bulletin. No. 137.
- [20] Jamil M., Iqbal W., Bangash A., Rehman S.U., Imran Q.M. and Rha E.S. 2010. Constitutive expression of OSC3H33, OSC3H50 and OSC3H37 genes in rice under salt stress. *Pak J. Bot.*, 42: 4003-4009.
- [21] Khadija M, Misratia, Mohd Razi Ismail, Md Abdul Hakim, Mohamed Hanafi Musa and Adam Puteh. 2013. Effect of salinity and alleviating role of gibberillic acid (GA_3) for improving the morphological, physiological and yield traits of rice varieties. *AJCS* 7(11):1682-1692.
- [22] Khalehi E., Arazani K., Moallemi N., and Barzegar M. 2012. Evaluation of chlorophyll content and chlorophyll fluorescence parameters and relationships between chlorophyll a, b and chlorophyll content index under water stress in *Olea europaea* cv. Dezful. *World Academy of Science, Engineering and Technology*, 68:1154-1157.
- [23] Khan, M.A. and Z. Abdullah, 2003. Salinity sodicity induced changes in reproductive physiology of rice (*Oryza sativa* L.) under dense soil conditions. *Environ. Exp. Bot.* 49: 145-157.
- [24] Kim .S. K., T. K. Son, S. Y. Park, I. J. Lee, B. H. Lee, H. Y. Kim and S. C. Lee. 2006. Influences of gibberellin and auxin on endogenous plant hormone and starch mobilization during rice seed germination under salt stress. *Journal of Environmental Biology*. 27(2) 181-186.
- [25] Linghe Zeng and Michael C. Shannon. 2000. Salinity effects on seedling growth and yield components of Rice. *Crop sci.* 40: 996-1003.
- [26] Mansour M.M. and Salama K.H. 2004. Cellular basis of salinity tolerance in plants. *Environ exp. Bot* 52: 113-122.
- [27] Mirza Hasamuzzaman, Masayuki Fujita , M.N. Islam, K.U. Ahamed , Kamrun Nahar. 2009. Performance of four irrigated rice varieties under different levels of salinity stress. *International Journal of Integrative Biology*: 6: 2-85.
- [28] Mohammad Reza Amirjan. 2011. Effect of Salinity Stress on Growth, Sugar content, Pigments and Enzyme activity of Rice. *International Journal of Botany*. 7(1): 73-81.
- [29] Munns, R. 2005. Genes and salt tolerance bringing them together. *New phytology*, 167: 645-663.
- [30] Murphy, K.S.T and M.J. Durako. 2003. Physiological effects of short term salinity changes on *Ruppia maritima*. *Aquatic Botany*. 75: 293-309.
- [31] Nawaz K., Talat A.I., Hussain K. and Majeed A. 2010. Induction of salt tolerance in two cultivars of sorghum (*Sorghum bicolor* L.) by exogenous application of proline at seedling stage. *World Applied Sciences Journal*, 10: 93-99.
- [32] Osakabe Y., Kajita S. and Osakabe K. 2011. Genetic engineering of woody plants: current and future targets in a stressful environment. *Physiol. Plant.* 142: 105-117.
- [33] Parvaneh Rahdari and Seyed Meysam Hoseini. 2015. Evaluation of Germination Percentage and Some Physiologic Factors under Salinity Stress and Gibberillic acid Hormone (GA_3) Treatments in Wheat (*Triticum aestivum* L.). *Int. J. Adv. Res. Biol.Sci.* 2(2): 122-131.
- [34] Plaut A., Edelstein M. Ben Hur . 2013. Overcoming salinity barriers to crop production using traditional methods. *Crit. Rev. Plant Sci.* 32(4): 250-291.
- [35] Rahdari, P. S. Tavakoli, and S.M. Hosseini, 2012. Studying stress effect on germination, Proline, Sugar, Protein, Lipid and Chlorophyll content in Purslane (*Portulaca oleracea* L.) leaves. *Journal of Stress Physiology & Biochemistry* 8(1): 182-193.
- [36] Rahman M., Soomro U.A., Haq M.Z., and Gul, S., 2008. Effects of NaCl salinity on wheat (*Triticum aestivum* L.) cultivars *World J. Agric. Sci.*, 4(3): 398-403.
- [37] Rahman M.S., H. Miyake and Y. Takeoka, 2001. Effect of sodium chloride salinity on seed germination and early seedling growth of Rice (*Oryza sativa* L.) *Pakistan Journal of Biological Sciences* 4(3): 351-355.
- [38] Razaque M. A., Talukder N.M., Islam M.S., Bhadra A.K. and Dutta R.K., 2009. The effect of salinity on morphological characteristics of seven rice (*Oryza sativa* L.) genotypes differing in salt tolerance. *Pakistan J. Biological Sci.*, 12(5):406-412.
- [39] Reddy, M.P. and A.B. Vora. 1986. Changes in pigment composition, hill reaction activity and saccharides metabolism in bajra (*Pennisetum typhoides* S&H) leaves under NaCl salinity. *Photosynthetica*, 20: 50-55.
- [40] Rou N.P., Tripathi S.B. and Shaw B.P. 1997. Effect of salinity on chlorophyll and proline contents in three aquatic macrophytes. *Biol. Plant.* 40: 453-458.
- [41] Roy, A.K.D., K. Alam and J. Gow. 2012. A review of the role of property rights and forest policies in the management of the Sundarbans mangrove forest in Bangladesh. *Forest Polciy Econ.* 15: 46-53.
- [42] Roychoudhury A., Basu S., Sengupta D.N. 2011. Amelioration of salinity stress by exogenously applied spermidine or spermine in three varieties of rice differing in their level of salt tolerance. *J. Plant Physiol.* 168(4): 317-328.
- [43] Sairam R.K. and A. Tyagi 2004. Physiology and molecular biology of salinity stress tolerance in plants. *Curr.Sci.*, 86:407-421.



- [44] Saleethong P, Sanitchon J, Kong-ngern K, Theerakulpisut P (2013) Effects of exogenous spermidine (Spd) on yield, yield-related parameters and mineral composition of rice (*Oryza sativa* L. ssp. indica) grains under salt stress. *Aust J Crop Sci* 7:1293-1301.
- [45] Santo, C.V. 2004. Regulation of chlorophyll biosynthesis and degradation by salt stress in sunflower leaves. *Science of Horticulure* 103, 93-99.
- [46] Seeman, J. R. and C. Critchley.1985. Effects of salt stress on the growth, ion content, stomatal behavior and photosynthetic capacity of a salt-sensitive species, *Phaseolus vulgaris* L. *Planta* 164:151-162.
- [47] Shannon M.C., Rhodes J.D., Draper J.H., Scardaci S.C., Spyres M.D., 1998. Assessment of salt tolerance in rice cultivars in response to salinity problems in California. *Crop Science* 38: 394-398.
- [48] Siringam, K., N. Juntawong, S. Chaum and C. Kirdmance 2011. Salt stress induced ion accumulation, ionhomeostasis, membrane injury and sugar contents in salt sensitive rice (*Oryza sativa* L. Spp. Indica) roots under isosmotic conditions. *Afr. J. Biotech.* 10: 1340-1346.
- [49] Thanaphol Boriboonkaset, Cattarin Theerawitaya, Aussanee Pichakum, Suriyan Cha-um, Teruhiro Takabe and Chalernpol Kirdmanee. 2012. Expression levels of some starch metabolism related genes in flag leaf of two contrasting rice genotypes exposed to salt stress. *AJCS* 6(11):1579-1586.
- [50] Yeo A.R. and T.J. Flowers. 1984. Mechanisms of salinity resistance in rice and their role as physiological criteria in plant breeding. P. 151-170. In: *Salinity tolerance in plants strategies for crop improvement* R.C. Staples and G.A. Toennisessen (eds). John Wiley and Sons. New York. USA.
- [51] Yeo A.R., K.S. Lee., P. Izard., P.J. Boursier and T.J. Flowers 1991. Short and long term effect of salinity on leaf growth in rice (*Oryza sativa* L.). *J. Exp. Bot.* 42:881-889.
- [52] Zeng, L. And Shannon, M.C. 2000b. Salinity effects on seedling growth and yield components of rice. *Crop Sci.*, 40:996-1003.
- [53] Zhu J.K. 2001. Plant salt tolerance. *Trends Plant Sci.* 6:66-71.



10.22214/IJRASET



45.98



IMPACT FACTOR:
7.129



IMPACT FACTOR:
7.429



INTERNATIONAL JOURNAL FOR RESEARCH

IN APPLIED SCIENCE & ENGINEERING TECHNOLOGY

Call : 08813907089  (24*7 Support on Whatsapp)